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FATTY ACIDS

(57) Abstract

By this invention, compositions and methods of use related to β -ketoacyl-ACP synthase of special interest are synthases obtainable from Cuphea species. Amino acid and nucleic acid for synthase protein factors are provided, as well as methods to utilize such sequences in constructs for production of genetically engineered plants having altered fatty acid compositions. Of particular interest is the expression of synthase protein factors in conjunction with expression of plant medium-chain acyl-ACP thioesterases for production of increased levels and/or modified ratios of medium-chain fatty acids in oils of transgenic plant seeds.

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PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

INTRODUCTION

Field of Invention

The present invention is directed to genes encoding plant fatty acid synthase enzymes relevant to fatty acid synthesis in plants, and to methods of using such genes in combination with genes encoding plant medium-chain preferring thioesterase proteins. Such uses provide a method to increase the levels of medium-chain fatty acids that may be produced in seed oils of transgenic plants.

Background

Higher plants synthesize fatty acids via a common metabolic pathway. In developing seeds, where fatty acids attached to triglycerides are stored as a source of energy for further germination, the fatty acid synthesis pathway is located in the plastids. The first step is the formation of acetyl-ACP (acyl carrier protein) from acetyl-CoA and ACP catalyzed by a short chain preferring condensing enzyme, ß-ketoacyl-ACP synthase (KAS) III. Elongation of acetyl-ACP to 16- and 18- carbon fatty acids involves the cyclical action of the following sequence of reactions: condensation with a two-carbon unit from malonyl-ACP to form a longer ß-ketoacyl-ACP (ß-ketoacyl-ACP synthase), reduction of the

keto-function to an alcohol (ß-ketoacyl-ACP reductase), dehydration to form an enoyl-ACP (ß-hydroxyacyl-ACP dehydrase), and finally reduction of the enoyl-ACP to form the elongated saturated acyl-ACP (enoyl-ACP reductase). ß-ketoacyl-ACP synthase I (KAS I), is primarily responsible for elongation up to palmitoyl-ACP (C16:0), whereas ß-ketoacyl-ACP synthase II (KAS II) is predominantly responsible for the final elongation to stearoyl-ACP (C18:0).

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Genes encoding peptide components of ß-ketoacyl-ACP synthases I and II have been cloned from a number of higher plant species, including castor (Ricinus communis) and Brassica species (USPN 5,510,255). KAS I activity was associated with a single synthase protein factor having an approximate molecular weight of 50 kD (synthase factor B) and KAS II activity was associated with a combination of two synthase protein factors, the 50 kD synthase factor B and a 46 kd protein designated synthase factor A. Cloning and sequence of a plant gene encoding a KAS III protein has been reported by Tai and Jaworski (Plant Physiol. (1993) 103:1361-1367).

The end products of plant fatty acid synthetase activities are usually 16- and 18-carbon fatty acids. There are, however, several plant families that store large amounts of 8- to 14-carbon (medium-chain) fatty acids in their oilseeds. Recent studies with Umbellularia californica (California bay), a plant that produces seed oil rich in lauric acid (12:0), have demonstrated the existence of a medium-chain-specific isozyme of acyl-ACP thioesterase

in the seed plastids. Subsequent purification of the 12:0-ACP thioesterase from *Umbellularia californica* led to the cloning of a thioesterase cDNA which was expressed in seeds of *Arabidopsis* and *Brassica* resulting in a substantial accumulation of lauric acid in the triglyceride pools of these transgenic seeds (USPN 5,512,482). These results and subsequent studies with medium-chain thioesterases from other plant species have confirmed the chain-length-determining role of acyl-ACP thioesterases during de novo fatty acid biosynthesis (T. Voelker (1996) *Genetic Engineering*, Ed. J. K. Setlow, Vol. 18, pgs. 111-133).

DESCRIPTION OF THE FIGURES

Figure 1. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-2 are provided. Figure 2. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-31-7 are provided. Figure 3. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor A clone chKAS A-2-7 are provided. Figure 4. DNA and translated amino acid sequence of Cuphea 20 hookeriana KAS factor A clone chKAS A-1-6 are provided. Figure 5. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/7-8 are provided. Figure 6. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/8-7A are provided. 25 Figure 7. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p7-6A are provided. Figure 8. Preliminary DNA sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p8-9A is provided.

- Figure 9. DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 are provided.
 - Figure 10. The activity profile for purified cpuKAS B/8-7A using various acyl-ACP substrates is provided.
- Figure 11. The activity profile for purified chKAS A-2-7 and chKAS A-1-6 using various acyl-ACP substrates is provided.
 - Figure 12. The activity profile for purified castor KAS factor A using various acyl-ACP substrates is provided.
- 10 Figure 13. The activity profile for purified castor KAS factor B using various acyl-ACP substrates is provided.

 Figure 14. A graph showing the number of plants arranged according to C8:0 content for transgenic plants containing CpFatB1 versus transgenic plants containing CpFatB1 + chKAS

A-2-7 is provided.

- Figure 15. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
- Figure 16. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
- Figure 17. Graphs showing the %C10/%C8 ratios in transgenic
 plants containing ChFatB2 (4804-22-357) and in plants
 resulting from crosses between 4804-22-357 and 5413-17 (chKAS
 A-2-7 + CpFatB1 plants) are provided.
 - Figure 18. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in

plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.

Figure 19. Graphs showing the %C12:0 in transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 20. Graph showing the relative proportions of C12:0 and C14:0 fatty acids in the seeds of transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 21. Graphs showing the %C18:0 in transgenic plants containing Garm FatB1 (5266) and in seeds of plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 22. The activity profile of Ch KAS A in protein extracts from transgenic plants containing Ch KAS A-2-7. Extracts were preptreated with the indicated concentrations of cerulenin.

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SUMMARY OF THE INVENTION

By this invention, compositions and methods of use related to ß-ketoacyl-ACP synthase (KAS) are provided. Also of interest are methods and compositions of amino acid and nucleic acid sequences related to biologically active plant synthase(s).

In particular, genes encoding KAS protein factors A and B from Cuphea species are provided. The KAS genes are of interest for use in a variety of applications, and may be

used to provide synthase I and/or synthase II activities in transformed host cells, including bacterial cells, such as E. coli, and plant cells. Synthase activities are distinguished by the preferential activity towards longer and shorter acyl-ACPs as well as by the sensitivity towards the KAS specific inhibitor, cerulenin. Synthase protein preparations having preferential activity towards medium chain length acyl-ACPs are synthase I-type or KAS I. The KAS I class is sensitive to inhibition by cerulenin at concentrations as low as $1\mu M$. Synthases having preferential activity towards longer chain length acyl-ACPs are synthase II-type or KAS II. The KAS enzymes of the II-type are also sensitive to cerulenin, but at higher concentrations (50 μ M). Synthase III-type enzymes have preferential activity towards short chain length acyl-ACPs and are insensitive to cerulenin inhibition.

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Nucleic acid sequences encoding a synthase protein may be employed in nucleic acid constructs to modulate the amount of synthase activity present in the host cell, especially the relative amounts of synthase I-type, synthase II-type and synthase III-type activity when the host cell is a plant host cell. In addition, nucleic acid constructs may be designed to decrease expression of endogenous synthase in a plant cell as well. One example is the use of an antisense synthase sequence under the control of a promoter capable of expression in at least those plant cells which normally produce the enzyme.

Of particular interest in the present invention is the coordinate expression of a synthase protein with the

expression of thioesterase proteins. For example, coordinated expression of synthase factor A and a medium-chain thioesterase provides a method for increasing the level of medium-chain fatty acids which may be harvested from transgenic plant seeds. Furthermore, coordinated expression of a synthase factor A gene with plant medium-chain thioesterase proteins also provides a method by which the ratios of various medium-chain fatty acids produced in a transgenic plant may be modified. For example, by expression of a synthase factor A, it is possible to increase the ratio of C10/C8 fatty acids which are produced in plant seed oils as the result of expression of a thioesterase having activity on C8 and C10 fatty acids.

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DETAILED DESCRIPTION OF THE INVENTION

A plant synthase factor protein of this invention includes a sequence of amino acids or polypeptide which is required for catalyzation of a condensation reaction between an acyl-ACP having a chain length of C2 to C16 and malonyl-ACP in a plant host cell. A particular plant synthase factor protein may be capable of catalyzing a synthase reaction in a plant host cell (for example as a monomer or homodimer) or may be one component of a multiple peptide enzyme which is capable of catalyzing a synthase reaction in a plant host cell, i.e. one peptide of a heterodimer.

Synthase I (KAS I) demonstrates preferential activity towards acyl-ACPs having shorter carbon chains, C2-C14 and is sensitive to inhibition by cerulenin at concentrations of 1µM. Synthase II (KAS II) demonstrates preferential

activity towards acyl-ACPs having longer carbon chains, C14-C16, and is inhibited by concentrations of cerulenin (50µM). Synthase III demonstrates preferential activity towards acyl-CoAs having very short carbon chains, C2 to C6, and is insensitive to inhibition by cerulenin.

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Synthase factors A and B, and synthase III proteins obtained from medium-chain fatty acid producing plant species of the genus Cuphea are described herein. As described in the following Examples, synthase A from C. hookeriana is naturally expressed at a high level and only in the seeds. C. hookeriana synthase B is expressed at low levels in all tissues examined. Expression of synthase A and synthase B factors in E. coli and purification of the resulting proteins is employed to determine activity of the various synthase factors. Results of these analyses indicate that synthase factor A from Cuphea hookeriana has the greatest activity on 6:0-ACP substrates, whereas synthase factor B from Cuphea pullcherrima has greatest activity on 14:0-ACP. Similar studies with synthase factors A and B from castor demonstrate similar activity profiles between the factor B synthase proteins from Cuphea and The synthase A clone from castor, however, castor. demonstrates a preference for 14:0-ACP substrate.

Expression of a Cuphea hookeriana KAS A protein in transgenic plant seeds which normally do not produce medium-chain fatty acids does not result in any detectable modification of the fatty acid types and contents produced in such seeds. However, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a

medium-chain acyl-ACP thioesterase capable of providing for production of C8 and C10 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are observed. In addition, where significant amounts of C8 and C10 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C8 and C10 fatty acids that are obtained. For example, an increased proportion of C10 fatty acids may be obtained by co-expression of Cuphea hookeriana ChFatB2 thioesterase and a chKAS A synthase factor proteins.

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Furthermore, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a medium-chain acyl-ACP thioesterase capable of providing for production of C12 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are also observed. In addition, where significant amounts of C12 and C14 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C12 and C14 fatty acids that are obtained. For example, an increased proportion of C12 fatty acids may be obtained by co-expression of UC FatB1 thioesterase and a chKAS A synthase factor proteins.

However, when Cuphea hookeriana KAS A protein is expressed in conjunction with the expression of a long-chain

acyl-ACP thioesterase capable of providing for production of C18 and C18:1 fatty acids in plant seed oils, no effect on the production of long chain fatty acids was observed.

Furthermore, when plants transformed to express a long chain acyl-ACP thioesterase from mangosteen (GarmFatA1, U.S. Patent Application No. 08/440,845), which preferentially hydrolyzes C18:0 and C18:1 fatty acyl-ACPs, are crossed with nontransformed control plants, a significant reduction in the levels of C18:0 is obtained. Similar reductions are also observed in the levels of C18:0 in the seeds of plants resulting from crosses between plants transformed to express the GarmFatA1 and plants expressing the Cuphea hookeriana KAS A protein.

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Thus, the instant invention provides methods of increasing and/or altering the medium-chain fatty acid compositions in transgenic plant seed oils by co-expression of medium-chain acyl-ACP thioesterases with synthase factor proteins. Furthermore, various combinations of synthase factors and medium-chain thioesterases may be achieved depending upon the particular fatty acids desired. For example, for increased production of C14 fatty acids, synthase protein factors may be expressed in combination with a C14 thioesterase, for example from Cuphea palustris or nutmeg may be employed (WO 96/23892). In addition, thioesterase expression may be combined with a number of different synthase factor proteins for additional effects on medium-chain fatty acid composition.

Synthases of use in the present invention include modified amino acid sequences, such as sequences which have

been mutated, truncated, increased and the like, as well as such sequences which are partially or wholly artificially synthesized. The synthase protein encoding sequences provided herein may be employed in probes for further screening or used in genetic engineering constructs for transcription or transcription and translation in host cells, especially plant host cells. One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and used to screen and recover synthases and/or synthase nucleic acid sequences from other sources. Typically, a homologously related nucleic acid sequence will show at least about 60% homology, and more preferably at least about 70% homology, between the R. communis synthase and the given plant synthase of interest, excluding any deletions which may be present. Homology is determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization reactions.

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Recombinant constructs containing a nucleic acid sequence encoding a synthase protein factor or nucleic acid sequences encoding a synthase protein factor and a medium-chain acyl-ACP thioesterase may be prepared by methods well known in the art. Constructs may be designed to produce synthase in either prokaryotic or eukaryotic cells. The increased expression of a synthase in a plant cell, particularly in conjunction with expression of medium-chain thioesterases, or decreasing the amount of endogenous synthase observed in plant cells are of special interest.

Synthase protein factors may be used, alone or in combination, to catalyze the elongating condensation reactions of fatty acid synthesis depending upon the desired result. For example, rate influencing synthase activity may reside in synthase I-type, synthase II-type, synthase III-type or in a combination of these enzymes. Furthermore, synthase activities may rely on a combination of the various synthase factors described herein.

Constructs which contain elements to provide the transcription and translation of a nucleic acid sequence of interest in a host cell are "expression cassettes".

Depending upon the host, the regulatory regions will vary, including regions from structural genes from viruses, plasmid or chromosomal genes, or the like. For expression in prokaryotic or eukaryotic microorganisms, particularly unicellular hosts, a wide variety of constitutive or regulatable promoters may be employed. Among transcriptional initiation regions which have been described are regions from bacterial and yeast hosts, such as E. coli, B. subtilis, Saccharomyces cerevisiae, including genes such as ß-galactosidase, T7 polymerase, trp-lac (tac), trp E and the like.

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An expression cassette for expression of synthase in a plant cell will include, in the 5' to 3' direction of transcription, a transcription and translation initiation control regulatory region (also known as a "promoter") functional in a plant cell, a nucleic acid sequence encoding a synthase, and a transcription termination region.

Numerous transcription initiation regions are available

which provide for a wide variety of constitutive or regulatable, e.g., inducible, transcription of the desaturase structural gene. Among transcriptional initiation regions used for plants are such regions associated with cauliflower mosaic viruses (35S, 19S), and structural genes such as for nopaline synthase or mannopine synthase or napin and ACP promoters, etc. The transcription/ translation initiation regions corresponding to such structural genes are found immediately 5' upstream to the respective start codons. Thus, depending upon the intended use, different promoters may be desired.

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Of special interest in this invention are the use of promoters which are capable of preferentially expressing the synthase in seed tissue, in particular, at early stages of seed oil formation. Examples of such seed-specific promoters include the region immediately 5' upstream of a napin or seed ACP genes such as described in USPN 5,420,034, desaturase genes such as described in Thompson et al (Proc. Nat. Acad. Sci. (1991) 88:2578-2582), or a Bce-4 gene such as described in USPN 5,530,194. Alternatively, the use of the 5' regulatory region associated with the plant synthase structural gene, i.e., the region immediately 5' upstream to a plant synthase structural gene and/or the transcription termination regions found immediately 3' downstream to the plant synthase structural gene, may often be desired. general, promoters will be selected based upon their expression profile which may change given the particular application.

In addition, one may choose to provide for the transcription or transcription and translation of one or more other sequences of interest in concert with the expression or anti-sense of the synthase sequence, particularly medium-chain plant thioesterases such as described in USPN 5,512,482, to affect alterations in the amounts and/or composition of plant oils.

When one wishes to provide a plant transformed for the combined effect of more than one nucleic acid sequence of interest, a separate nucleic acid construct may be provided for each or the constructs may both be present on the same plant transformation construct. The constructs may be introduced into the host cells by the same or different methods, including the introduction of such a trait by crossing transgenic plants via traditional plant breeding methods, so long as the resulting product is a plant having both characteristics integrated into its genome.

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Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformed cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an auxotrophic host, viral immunity or the like. Depending upon the number of different host species into which the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

The manner in which the DNA construct is introduced into the plant host is not critical to this invention. Any method which provides for efficient transformation may be employed. Various methods for plant cell transformation include the use of Ti- or Ri-plasmids, microinjection, electroporation, liposome fusion, DNA bombardment or the like. In many instances, it will be desirable to have the construct bordered on one or both sides by T-DNA, particularly having the left and right borders, more particularly the right border. This is particularly useful when the construct uses A. tumefaciens or A. rhizogenes as a mode for transformation, although the T-DNA borders may find use with other modes of transformation.

The expression constructs may be employed with a wide variety of plant life, particularly plant life involved in the production of vegetable oils. These plants include, but are not limited to rapeseed, peanut, sunflower, safflower, cotton, soybean, corn and oilseed palm.

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For transformation of plant cells using Agrobacterium, explants may be combined and incubated with the transformed Agrobacterium for sufficient time for transformation, the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

EXAMPLES

Example 1 Cuphea KAS Factor A and B Gene Cloning

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Total RNA isolated from developing seeds of Cuphea hookeriana and Cuphea pullcherrima was used for cDNA synthesis in commercial 1-based cloning vectors. For cloning each type of KAS gene, approximately 400,000-500,000 unamplified recombinant phage were plated and the plaques transferred to nitrocellulose. For KAS factor B cloning from C. hookeriana, a mixed probe containing Brassica napus KAS factor B and Ricinus communis (Castor) KAS factor B radiolabeled cDNA's was used. Similarly, a mixed probe containing Brassica napus KAS factor A and Ricinus communis KAS factor A cDNA clones was used to obtain C. hookeriana KAS factor A genes. For KASIII, a spinach KASIII cDNA clone obtained from Dr. Jan Jaworski was radiolabeled and used as a probe to isolate a KASIII clone from C. hookeriana. For KAS B and KAS A cloning from C. pullcherrima, C. hookeriana KAS B and KAS A genes chKAS B-2 and chKAS A-2-7 (see below) were radiolabeled and used as probes.

DNA sequence and translated amino acid sequence for Cuphea KAS clones are provided in Figures 1-9. Cuphea hookeriana KAS factor B clones chKAS B-2 and chKAS B-31-7

are provided in Figures 1 and 2. Neither of the clones is full length. Cuphea hookeriana KAS Factor A clones chKAS A-2-7 and chKAS A-1-6 are provided in Figures 3 and 4. chKAS A-2-7 contains the entire encoding sequence for the KAS factor protein. Based on comparison with other plant synthase proteins, the transit peptide is believed to be represented in the amino acids encoded by nucleotides 125-466. chKAS A-1-6 is not a full length clone although some transit peptide encoding sequence is present. Nucleotides 1-180 represent transit peptide encoding sequence, and the mature protein encoding sequence is believed to begin at nucleotide 181.

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Cuphea pullcherrima KAS factor B clones cpuKAS B/7-8 and cpuKAS B/8-7A are provided in Figures 5 and 6. Both of the clones contain the entire encoding sequences for the KAS factor B proteins. The first 35 amino acids of cpuKAS B/7-8 are believed to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide The first 39 amino acids of cpuKAS B/8-7A are believed 233. to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide 209. Cuphea pullcherrima KAS factor A clones cpuKAS A/p7-6A and cpuKAS A-p8-9A are provided in Figures 7 and 8. Both of the clones contain the entire encoding sequences for the KAS factor A proteins. Translated amino acid sequence of cpuKAS A/p7-6A The mature protein is believed to begin at the is provided. lysine residue encoded 595-597, and the first 126 amino acids are believed to represent the transit peptide. DNA sequence of KAS A clone cpuKAS A-p8-9A is preliminary.

Further analysis will be conducted to determine final DNA sequence and reveal the amino acid sequence encoded by this gene.

DNA and translated amino acid sequence of *Cuphea hookeriana* KASIII clone chKASIII-27 is provided in Figure 9. The encoding sequence from nucleotides 37-144 of chKASIII-27 are believed to encode a transit peptide, and the presumed mature protein encoding sequence is from nucleotides 145-1233.

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Deduced amino acid sequence of the C. hookeriana KAS factor B and KAS factor A cDNA's reveals strong homology to the Brassica napus and Ricinus communis clones previously reported. The C. hookeriana KAS factor B clone is more homologous to the Ricinus and Brassica KAS factor B clones (94% and 91% respectively) than it is to the Ricinus and Brassica KAS factor A clones (60% for both). Furthermore, the C. hookeriana KAS factor A clone is more homologous to the Ricinus and Brassica KAS factor A clones (85% and 82% respectively) than it is the Ricinus and Brassica KAS factor B clone (60% for both). The C. hookeriana KAS factor B cDNAs designated as chKAS B-2 and chKAS B-31-7 are 96% identical within the mature portion of the polypeptide. Similarly, the deduced amino acid sequence of the mature protein regions of the C. hookeriana KAS factor A clones chKAS A-2-7 and chKAS A-1-6 are 96% identical. pullcherrima KAS clones also demonstrate homology to the R. communis and Brassica napus KAS clones. The mature protein portion of all of the KAS factor A family members in the different Cuphea species are 95% identical. Similarly the

mature protein portion of the KAS factor B genes in Cuphea are also 95-97% identical with each other. However there is only approximately 60% sequence identity between KAS factor B and KAS factor A clones either within the same or different species of Cuphea.

Example 2 Levels and Patterns of Expression

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To examine tissue specificity of KAS expression in Cuphea hookeriana, Northern blot analysis was conducted using total RNA isolated from seed, root, leaf and flower tissue. Two separate but identical blots were hybridized with either chKAS B-31-7 or chKAS A-2-7 coding region probes. The data from this RNA blot analysis indicate that KAS B is expressed at a similar level in all tissues examined, whereas KAS A expression is detected only in the These results also demonstrate a different level of seed. expression for each of the synthases. KAS A is an abundant message, whereas KAS B is expressed at low levels. Furthermore, even under highly stringent hybridization conditions (65_C, 0.1 X SSC, 0.5% SDS), the KAS A probe hybridizes equally well with two seed transcripts of 2.3 and The larger hybridizing band is likely the 1.9 kb. transcript of the KAS A-2-7 gene since the size of its cDNA is 2046bp, and the number of clones obtained from cDNA screening corresponds well with the apparent mobility of the mRNA and its abundance on the blot.

Example 3 Expression of Plant KAS Genes in E.coli

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DNA fragments encoding the mature polypeptide of the Cuphea hookeriana KAS A cDNAs and the Cuphea pullcherrima 5 KAS B cDNAs were obtained by PCR and cloned into a QIAexpress expression vector (Qiagene). Experimental conditions for maximum level of expression were determined for all of these clones and the parameters for highest level of soluble fraction were identified. Cells are grown in ECLB media containing 1M sorbitol and 2.5 mM betaine overnight and subcultured as a 1:4 dilution in the same medium. Cells are then grown for 2 hours (to approximately .6-.8 O.D.) and induced with 0.4 mM IPTG and allowed to grow for 5 more hours.

Enzyme activity of the affinity purified recombinant enzymes obtained from over-expression of the chKAS A-2-7 and cpuKAS B/8-7A clones was measured using a wide range of acyl-ACP substrates (6:0- to 16:1-ACP). The activity profile for cpuKAS B/8-7A is provided in Fig.10. demonstrate that the enzyme is active with all acyl-ACP substrates examined, although activity on 6:0 to 14:0-ACP substrates is substantially greater than the activity on 16:0 and 16:1 substrates.

The activity profile of the C. hookeriana KAS A clones chKAS A-2-7 and chKAS A-1-6 is provided in Figure 11. The C. 25 hookeriana KAS A clones are most active with C:6, and have the least activity with C:16:0 substrates. However, the activity of this clone on even the preferred C6:0 substrate

is 50 fold lower than the activity of the *C. pullcherrima* KAS B clones.

A fragment containing the mature protein encoding portion of a R. communis KAS factor A clone was also cloned into a QIAexpress expression vector, expressed in E. coli and the enzyme affinity purified as described above. The activity profile for castor KAS A is provided in Figure 12. Highest activity is observed with C14:0 substrates, although some activity is also seen with C6:0 and C16:1. In comparison, the activity profile obtained from purified R. communis KAS factor B also using the QIAexpress expression system is provided in Figure 13. The KAS B clone demonstrates substantially higher levels of activity (10 fold and higher) than the R. communis KAS A clone. The preference of the KAS factor B for 6:0- to 14:0-ACP substrates is consistent with the previous observations that this protein provides KAS I activity.

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Example 4 KAS and TE Expression in Transgenic Seed

Both the CpFatB1 (*C. hookeriana* thioesterase cDNA;

Dehesh et al. (1996) Plant Physiol. 110:203-210) and the chKAS A-2-7 were PCR amplified, sequenced, and cloned into a napin expression cassette. The napin/cp FatB1 and the napin/KAS A-2-7 fusions were ligated separately into the binary vector pCGN1558 (McBride and Summerfelt (Pl.Mol.Biol. (1990) 14:269-276) and transformed into A. tumefaciens, EHA101. The resulting CpFatB1 binary construct is pCGN5400 and the chKAS A-2-7 construct is pCGN5401. Agrobacterium mediated transformation of a Brassica napus canola variety

was carried out as described by Radke et al. (Theor. Appl. Genet. (1988) 75:685-694; Plant Cell Reports (1992) 11:499-505). Several transgenic events were produced for each of the pCGN5400 and pCGN5401 constructs.

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A double gene construct containing a napin/cpFatB1 expression construct in combination with a napin/chKAS A-2-7 expression construct was also assembled, ligated into a binary vector and used for co-cultivation of a canola Brassica variety. The binary construct containing the chFatB1 and chKAS A-2-7 expression constructs is pCGN5413.

Fatty acid analysis of 26 transgenic lines containing chKAS A-2-7 (5401 lines) showed no significant changes in the oil content or profile as compared to similar analyses of wild type canola seeds of the transformed variety.

Fatty acid analysis of 36 transgenic lines containing cpFatB1 (5400 lines) showed increased levels of C:8 and C:10 in transgenic seeds. The highest level of C:8 observed in a pool seed sample was 4.2 mol%. The C:10 levels were between 30 and 35% of the C:8 content. Fatty acid analysis of 25 transgenic lines containing the TE/KAS A tandem (5413 lines) demonstrated an overall increase in both C:8 and C:10 levels relative to those observed with TE containing lines (5400) alone. In lines containing the cpFatB1 construct alone, the average level of C:8 average were 1.5 mol%, whereas the C:8 average levels in TE/KAS A tandem containing lines was 2.37 mol%. The ratio of C:8 to C:10 remained constant in both populations. The number of transgenic events relative to the C:8 content are presented in Figure 14. These data show that the transgenic events with tandem TE/KAS A construct

yield more lines with higher levels of C:8 than those events with single TE construct. For example, several lines containing nearly 7 mole% C8 were obtained with the TE/KAS A pCGN5413 construct, whereas the highest C8 containing line from the pCGN5400 TE alone transformation contained 4.2 mole% C8.

Half seed analysis of the T3 generation of transgenic canola plants expressing a ChFatB2 (*C. hookeriana* thioesterase; Dehesh et al. (1996) The Plant Journal 9:167-172) indicate that these plant can accumulate up to 22 weight% (33 mol%) of 8:0 and 10:0 fatty acids (4804-22-357). Segregation analysis shows that these transformants contain two loci and that they are now homozygous. Selected plants grown from these half seeds were transferred into the greenhouse and later crossed with T1 transformants that had been transformed with either Cuphea hookeriana KAS A (5401) alone or KAS A/CpFatB1 double constructs (5413).

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Fatty acid analysis of several events resulting from the crosses between transgenic lines containing ChFatB2 (4804-22-357) and chKAS A-2-7 (5401-9), reveal an increase in the ratio of C:10/C:8 levels (Figure 15). This C:10/C:8 ratio in nearly all of the transgenic events containing ChFatB2 TE alone fluctuates between 3 and 6, whereas in the F1 generation of transgenic containing both the TE and the KAS A-2-7, the ratio can be as high as 22. This increase in C:10 levels is accompanied by an increase in the total C:8 and C:10 content (Figure 16). The sum of the C:8 and C:10 fatty acids in the heterozygous F1 lines is as high as those in the homozygous parent line (4804-22-357), whereas the

heterozygous lines usually contain substantially less C:8 and C:10 than the homozygous lines.

Similar results were observed in F1 generation seeds resulting from crosses performed between 4804-22-357 (ChFatB2) and the 5413-17 event (CpFatB1 and chKAS A-2-7 tandem). Levels of C:8 and C:10 in the 5413-17 line were 6.3 and 2.8 mol% respectively. Data presented in Figure 17 show that there is shift towards C:10 fatty acids as was observed with the 4804-22-357 (ChFatB2) x 5401-9 (chKAS A-2-7) crosses. Furthermore, Figure 18 indicates the presence 10 of two separate populations of heterozygotes. containing approximately 9-11 weight percent C:10 + C:8 are believed to represent offspring containing a single copy of the ChFatB1 TE gene and no copies of the CpFatB1 and chKAS A genes from 5413. Those plants containing approximately 15-15 20 weight percent C:10 + C:8 are believed to represent the heterozygotes containing a single ChFatB1 TE gene as well as the CpFatB1 and chKAS A genes from 5413. Thus, the level of the C:10 + C:8 fatty acids does not decrease to 50% of that detected in parent lines when a copy of the ChKAS A gene is 20 present.

To further characterize the chain length specificity of the Cuphea hookeriana KAS A enzyme, crosses between transgenic Brassica napus lines containing a California Bay (Umbellularia californica) 12:0 specific thioesterase, Uc FatB1 (USPN 5,344,771) and chKAS A-2-7 (5401-9) were made. Half seed analysis of transgenic plants containing Uc fatB1 have previuosly indicated that these plants can accumulate up to 52 mol% C12:0 in the seed oil of homozygous dihaploid

lines (LA86DH186). Crosses between the line LA86DH186 and untransformed control *Brassica* demonstrated a decrease in the C12:0 levels.

however, crosses between LA86DH186 and the 5401-9

hemizygous line led to an accumulation of up to 57 mol%

C12:0 in the seed oil of F1 progeny (Figure 19).

Interestingly, in crosses with LA86DH186 x untransformed control line and LA86DH186 x 5401-9, levels of C14:0 in the seeds of the F1 progeny decreased to 50% of the levels

obtained in homozygous LA86DH186 lines (Figure 20).

Furthermore, increases in the proportion of C12:0 fatty acid resulted in a substantial decline in the proportions of all the long-chain fatty acyl groups (C16:0, C18:0, C18:2, and C18:3). These results indicate that the ChKAS A-2-7 is an enzyme with substrate specificity ranging from C6:0 to C10:0-ACP, and that its over-expression ultimately reduces the longer chain acyl-ACP pools.

Further evidence is obtained in support of the chain length specificity of the ChKAS A-2-7 in crosses of the 5401-9 line with a transgenic line (5266) expressing an 18:1/18:0 TE from Garcinia mangostana (GarmFatAl, US patent application No. 08/440,845). Transgenic Brassica line 5266 has been shown to accumulate up to 24 mol% C18:0 in the seed oil of homozygous lines (Figure 21). However, in the seed oil of F1 progeny of crosses between 5266 and 5401-9 levels of C18:0 were reduced to approximately 12 mol%. Furthermore, levels of C16:0 generated from these crosses was similar to the levels obtained from the seed oil of nontransgenic control plants.

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Example 5 In vitro Analysis of Plant KAS Enzymes

Seed extracts were prepared from developing seeds of nontransgenic controls or transgenic Brassica expressing chKAS A-2-7 as described in Slabaugh et al. (Plant Journal, 1998 in press) and Leonard et al. (Plant Journal, 1998, in press). In vitro fatty acid synthesis assays were performed as described by Post-Beittenmiller (J. Biol. Chem. (1991), 266:1858-1865). Extracts were concentrated by ammonium sulfate precipitation and desalting using P-6 columns (Bio-Rad, Hercules, CA). Reactions (65µ1) contained 0.1M 10 Tris/HCl (pH 8.0), 1 mM dithiothreitol, 25 mM recombinant spinach ACP1, 1 mM NADH, 2 mM NADPH, 50 µM malonyl-CoA, 10 μM [1-14C]acetyl-CoA (50 mCi/mmol), 1mg/ml BSA, and 0.25 mg/ml seed protein. Selected seed extracts were preincubated with cerulenin at 23°C for 10 min. Reaction 15 products were separated on an 18% acrlamide gel containing 2.25M urea, electroblotted onto to nitrocellulose and quntitated by phosporimaging using Image QuaNT software (Molecular Dynamics, Sunnyvale, CA). Authentic acyl-ACPs were run in parallel, immunoblotted and finally detected by 20 anti-ACP serum to confirm fatty acid chain lengths.

The results (Figure 22) indicate that the fatty acid synthesis capabilities of transgenic Brasica (5401-9) seed extracts was greater than that obtained from in the nontransgenic controls as measured by the relative abundance of C8:0- and C10:0-ACP at all time points tested. In addition, pretreatment of the extracts with cerulenin, markedly reduced the synthesis of longer chain fatty acids in both the transgenic and nontransgenic control seed

extracts. However, the extension of the spinach-ACP was much less inhibited in the seed extracts from the transgenic lines than in the seed extracts of nontransgenic control Brassica.

These data further support that Ch KAS A-2-7 is a condensing enzyme active on medium chain acyl-ACPs, and that expression of this enzyme in plants results in enlarged substrate pools to be hydrolyzed by medium-chain specific thioesterases. Furthermore, these data suggest that chKAS A-2-7 also is a cerulenin-resistant condensing enzyme.

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All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claim.

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MISSING UPON TIME OF PUBLICATION

- 13. The construct of Claim 5 wherein said encoding sequence is cpuKAS A/p8-9A.
- 14. The construct of Claim 5 wherein said encoding sequence is chKASIII-27.
- 15. An improved method for producing medium-chain fatty acids in transgenic plant seeds by expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant,

the improvement comprising expression of a plant synthase factor protein heterologous to said transgenic plant in conjunction with expression of said plant medium-chain thioesterase, whereby the percentage of medium-chain fatty acids produced in seeds expressing both a plant synthase factor protein and a plant medium-chain thioesterase protein is increased as compared to the percentage of medium-chain fatty acids produced in seeds expressing only said plant medium-chain thioesterase protein.

- 16. The method of Claim 15 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 20 17. The method of Claim 15 wherein said medium-chain thioesterase protein is a CpFatB1 protein.
 - 18. The method of Claim 15 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
- 19. The method of Claim 15 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
 - 20. The method of Claim 19 wherein said synthase factor A protein is from a Cuphea species.

- 21. The method of Claim 20 wherein said Cuphea species is C. hookeriana or C. pullcherrima.
 - 22. A method of altering the medium-chain fatty acid composition in plant seeds expressing a heterologous plant medium-chain preferring thioesterase, wherein said method comprises

providing for expression of a plant synthase factor protein heterologous to said transgenic plant in conjunction with expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant, whereby the composition of medium-chain fatty acids produced in said seeds is modified as compared to the composition of medium-chain fatty acids produced in seeds expressing said plant medium-chain thioesterase protein in the absence of expression of said plant synthase factor protein.

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- 23. The method of Claim 22 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 24. The method of Claim 22 wherein said medium-chain thioesterase protein is a CpFatBl protein.
- 25. The method of Claim 22 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
 - 26. The method of Claim 22 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
 - 27. The method of Claim 26 wherein said synthase factor A protein is from a Cuphea species.
 - 28. The method of Claim 27 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

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29. The method of Claim 22 wherein said fatty acid composition is enriched for C10 fatty acids.

- 30. The method of Claim 22 wherein said fatty acid composition is enriched for C12 fatty acids.
- 31. The method of Claim 22 wherein said fatty acid composition is enriched for at least one medium chain fatty acid and at least one other medium chain fatty acid is decreased.
- 32. The method of Claim 31 wherein said enriched fatty acid is C12 and said decreased fatty acid is C14.

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48	96	144	192	240	288	336	384
GGC	AAG Lys	GGT Gly	CAC His	GGG Gly	TCA Ser	GCT Ala	ACT Thr
CCG G	TCC A	GGT (GGT (Gly)	ATG	TAT Tyr	GCC Ala	66C 61y
CCC (CTC	ATG	AAG Lys	AAC Asn	AAC Asn	GCT Ala	GGA G1y
GAT Asp	CGC Arg	GGA Gly	GAG Glu	ACA Thr	CCA Pro	CAT	GCT
GTG Val	GAC Asp	ACA Thr	ATC Ile	ATT Ile	GGC Gly	TTC	ATT Ile
CTA Leu	GCC Ala	GGA Gly	CTT Leu	GCC Ala	ATG Met	TGC	ATG
GAA Glu	GGT Gly	GTC Val	TCT Ser	$\mathtt{TAT}\\\mathtt{TY}_{\mathcal{I}}$	CTC	TAC	CTT
CTA Leu	CTC Leu	CTG Leu	CAG Gln	CCC Pro	GGT Gly	AAC Asn	GAT
GCT Ala	GAT Asp	GTG Val	GTT Val	ATC Ile	TTT Phe	TCC Ser	GCT
GCC Ala	GCC Ala	GGA Gly	666 61y	TTC Phe	GAA Glu	ACT Thr	GAG
GCG Ala	CGA Arg	GCC Ala	GAC Asp	TTC	ATC Ile	GCC	GGT Gly
GTG Val	GCA Ala	AGA Arg	TCT Ser	CCT	GCT	TGT Cys	CGT
GCG Ala	TCG Ser	GAG Glu	TTC Phe	ACC Thr	CTC	GCA	CGC
ACC Thr	AAT Asn	AAG Lys	GTC Val	ATC Ile	CTG	ACT Thr	ATC Ile
TCC	AGG Arg	GAC Asp	ACT Thr	AAA Lys	GCC	TCC	CAT
AGC	TGC	ATC Ile	CTG	CGG	TCT Ser	ATT Ile	AAT Asn

IGURE 1

			,		•		
480	528	576	624	672	720	768	
IGG Irp	TTG Leu	ATT Ile	ACT Thr	AGC	GCT	ATC Ile	
			ATG Met	AGT	AAT	GCC	
	GGA Gly	CCG	CAC	GAG Glu	ATA Ile	AAT Asn	
	GCT Ala	GCA Ala	TAT Tyr	ATT Ile			
	GG'T Gly	GGA Gly	GCT Ala	TGC			
	GAA Glu	CGA	GAT Asp	TCT Ser			
CAG Gln	GGT Gly	AGA Arg	TGT Cys	TCT Ser		CTC Leu URE 1	
CCG Pro	ATG Met	ATG Met	AAC Asn	GTC			
	GTG Val	GCA Ala	ATC Ile	GGT Gly		666 61y	
GAT Asp	TTT Phe	CAT His	GCA Ala	CTT Leu			
AAC Asn	GGT	GAA Glu	GGT Gly				
AGG Arg	GAT Asp	TTG	GGA Gly				
CAA	CGT Arg	AGC Ser					
TCT	GAC Asp	GAG Glu					
TTG Leu	AAA Lys	ATG Met	GAG				
GCT	GAT Asp	GTG Val	GCA Ala	GAT Asp	CTT Leu	CAT	
	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu ATG GAG AGC TTG GAA CAT GCA ATG AGA CGA GGA GCA CCG ATT ATT Met Glu Ser Leu Glu His Ala Met Arg Arg Gly Ala Pro Ile Ile	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu ATG GAG AGC TTG GAA CAT GCA ATG AGA CGA GGA GCA CCG ATT ATT Met Glu Ser Leu Glu His Ala Met Arg Arg Gly Ala Pro Ile Ile GAG TAT TTG GGA GGT GCA ATC AAC TGT GAT GCT TAT CAC ATG ACT Glu Tyr Leu Gly Gly Ala Ile Asn Cys Asp Ala Tyr His Met Thr	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu ATG GAG AGC TTG GAA CAT GCA ATG AGA CGA GGA GCA CCG ATT ATT Met Glu Ser Leu Glu His Ala Met Arg Arg Gly Ala Pro Ile Ile GAG TAT TTG GGA GGT GCA ATC AAC TGT GAT GCT TAT CAC ATG ACT Glu Tyr Leu Gly Gly Ala Ile Asn Cys Asp Ala Tyr His Met Thr CCA AGG GCT GAT GGT CTT GGT GTC TCT TCT TGC ATT GAG AGT AGC Pro Arg Ala Asp Gly Leu Gly Val Ser Ser Cys Ile Glu Ser Ser	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GCT TTT GTG ATG GGT GAA GGT GCT GCA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu ATG GAG AGC TTG GAA CAT GCA ATG AGA CGA GGA GCA CCG ATT ATT Met Glu Ser Leu Glu His Ala Met Arg Arg Gly Ala Pro Ile Ile Glu Tyr Leu Gly Gly Ala Ile Asn Cys Asp Ala Tyr His Met Thr CCA AGG GCT GAT GTT GGT GTC TCT TCT TGC ATT GAG AGT AGC Pro Arg Ala Asp Gly Leu Gly Val Ser Ser Cys Ile Glu Ser Ser Glu Asp Ala Gly Val Ser Pro Glu Glu Val Asn Tyr Ile Asn Ala Glu Asp Ala Gly Val Ser Pro Glu Glu Val Asn Tyr Ile Asn Ala	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu GLG TAT TTG GAA CAT GCA ATG AGG GGA GCA CCG ATT ATT Met Glu Ser Leu Glu His Ala Met Arg Arg Gly Ala Pro Ile Ile GLU Tyr Leu Gly Gly Ala Ile Asn Cys Asp Ala Tyr His Met Thr GLA AGG GCT GAT GGT CTT GGT GTC TCT TCT TGC ATT GAG AGT AGC Pro Arg Ala Asp Gly Leu Gly Val Ser Ser Cys Ile Glu Ser Ser GLU Asp Ala Gly Val Ser Pro Glu Glu Val Asn Tyr Ile Asn Ala GCG ACT TCT ACT CTA GCT GGG GAT CTC GCC GAG ATA AAT GCT Ala Asp Ala Gly Val Ser Pro Glu Glu Val Asn Tyr Ile Asn Ala HIGURE I 2005

816	864	912	096	1008	1056	1116	1176	
AAG AAG GTT TTC AAG AAC ACA AAG GAT ATC AAA ATT AAT GCA ACT AAG Lys Lys Val Phe Lys Asn Thr Lys Asp Ile Lys Ile Asn Ala Thr Lys	TCA ATG ATC GGA CAC TGT CTT GGA GCA TCT GGA GGT CTT GAA GCT ATA Ser Met Ile Gly His Cys Leu Gly Ala Ser Gly Gly Leu Glu Ala Ile	GCG ACT ATT AAG GGA ATA AAC ACC GGC TGG CTT CAT CCC AGC ATT AAT Ala Thr Ile Lys Gly Ile Asn Thr Gly Trp Leu His Pro Ser Ile Asn	CAA TTC AAT CCT GAG CCA TCG GTG GAG TTC GAC ACT GTT GCC AAC AAG Gln Phe Asn Pro Glu Pro Ser Val Glu Phe Asp Thr Val Ala Asn Lys	AAG CAG CAA CAC GAA GTT AAC GTT GCG ATC TCG AAT TCA TTC GGA TTT Lys Gln Gln His Glu Val Asn Val Ala Ile Ser Asn Ser Phe Gly Phe	GGA GGC CAC AAC TCA GTC GTG GCT TTC TCG GCT TTC AAG CCA TGATTA Gly Gly His Asn Ser Val Val Ala Phe Ser Ala Phe Lys Pro	CCCATTTCAC AAGGTACTTG TCATTGAGAA TACGGATTAT GGACTTGCAG AGTAATTTCC	CCATGTTTGT CGGAAGAGCA TATTACCACG GTTGTCCGTC AAACCCATTT AGGATACTGT	

FIGURE 1 3 OF 4

1236	1296	1348
TCTATGTAAT AAAACTAAGG ATTATTAATT TCCCTTTTAA TCCTGTCTCC AGTTTGAGCA	TGAAATTATA TTTATTTTAT CTTAGAAAGG TCAAATAAGA TTTTGTTTTA CCTCTGTAAA	AA
TCCTGTCTCC	TTTTGTTTTA	AAAAAAAA
TCCCTTTTAA	TCAAATAAGA	TCTCAAAAAA
ATTATTAATT	CTTAGAAAGG	GGAAGTGCCG
AAAACTAAGG	TTTATTTAT	GTATTGGAAA
TCTATGTAAT	TGAAATTATA	ACTITIGITI GIAITGGAAA GGAAGIGCCG ICTCAAAAAA AAAAAAAAA AA

Sequence Range: 1 to 1704

	40 GTG Val>		GCA Ala>		TCT Ser>	190	GAC Asp>	240	CGG Arg>	CTC Leu>	•	GAA Glu>
	GNG Xxx		TCG Ser	140	GAC Asp	17	ATC Ile		ATC Ile	AGG Arg		CTC
	ACC Thr	90	AAT Asn	1	GTC Val		TTA Leu		CAG Gln	280 GAC AGG ASP Arg	330	GCT Ala
Ċ	30 TCC Ser		AGG Arg		GAC Asp		AGC Ser	230	GGC Gly	28 GAC Asp		AAG Lys
	AGC Ser		TGC Cys	0	TCC Ser	180	ATC Ile	(4	${\tt GGC}$	AAC Asn		AAG Lys
	TGG Trp	80	GGC Gly	130	GGC Gly		666 61y		TTC Phe	AAG Lys	320	GGG
6	20 AGC Ser		CCG		TTC Phe		AGC Ser	220	AGG Arg	270 GGG G1Y	,	GCC Ala
	AAA Lys		CCC Pro		GTA Val	1.70	GAG Glu	22	ACC Thr	GAC Asp		GTC Val
	AAC Asn	7.0	GAT Asp	120	TCC Ser	П	GGC Gly		CCC Pro	ATC Ile	310	ATT Ile
,		7	GTG Val		GTC Val		TCC Ser		TTC	260 TAC TYr	, m	TGC
•	10 AAA GGG Lys Gly		CTA		CTC	160	CTC	210	AAG Lys	2 GGA G1Y		TAC
	ACT Thr		GAA Glu	110	GGC Gly	16	CTC		TCC Ser	ACG Thr		CGC Arg
	CTC	09	CTA Leu	П	ATG Met		AAG Lys		GCT	250 C GCG n Ala	300	CTC
	ACC Thr		GCT Ala		GGC Gly		GAA Glu	200	GAC Asp	25 AAC Asn		TGC Cys
	TTA Leu		GCC Ala	0	GCC Ala	150	TAC Tyr	(4	TTC Phe	TTC Phe		GAT Asp
	AAA Lys	20	GCG Ala	100	CGA Arg		\mathtt{TAT}		CGC Arg	GGA G1y	90	GAC Asp

FIGURE 2 1/5

				•								
	AGA Arg>	0	TCT Ser>	480	CCG Pro>	GCC Ala>		TGT Cys>		CGA Arg>	. 0	ATT Ile>
380	GAG Glu	43	TTC		TCC	CTT Leu		GCA Ala	. 029	CGC Arg	670	ATC Ile
ς,	AAG Lys		GTC Val		AAG ATC Lys Ile	20 CTG Leu	570	ACT Thr	v	ATC Ile		GCA Ala
	GAT Asp		AĆC Thr	470		520 GCT C		TCA		CAT His		GCT
0	ATT Ile	420	CTA	7	CGG Arg	TCT Ser		ATT Ile	610	AAT	* 099	GAG Glu
.370	AAG Lys		GGC Gly		CAC His	GGG Gly	260	TCG	9	GCC		ACT Thr
	TCC		$_{\rm GLy}^{\rm GGT}$	460	$_{\rm GLY}^{\rm GGT}$	510 ATG Met	۵,	$\mathtt{T}\mathtt{A}\mathtt{T}$		GCT Ala		GGA Gly
	CTC	410	ATG Met	46	AAA Lys	AAC Asn		AAC Asn		GCC Ala	029	GGA G1y
360	AGC Ser	7	$_{\rm GLY}^{\rm GGT}$		GAG Glu	ACA Thr	250	CCA	* 009	$\mathtt{TAT}\\ \mathtt{TYF}$		GCT Ala
	GAA Glu		ACT Thr		ATC Ile	500 ATT Ile	7.	GGC Gly		TTT Phe		ATT Ile
	$_{\rm GGT}$	0	$_{\rm GLy}^{\rm GGA}$	450	CTC	GCC Ala		ATG Met		TGC Cys	640	ATG Met
350	GGC Gly	400	GTT Val		AAT Asn	TAT Tyr		CTG	290	TAC	9	CTC
m	CTC		CTA		CAG Gln	490 T CCC e Pro	540	$_{\rm GLY}^{\rm GGT}$	٠	AAC Asn		GAC Asp
•	GAT Asp		GTG Val	440	GTT Val	49 ATT Ile		TTG Leu		TCC		GCT
0	TCC	390	GGA Gly	7	GGG Gly	TTC Phe		GAT Asp	280	ACT Thr	630	GAG Glu
340	AAT Asn		GCT Ala		GAC Asp	TTT Phe	30	ATC Ile	25	GCT Ala		GGC Gly
							•					

SUBSTITUTE SHEET (RULE 26)

FIGURE 2 3/5

720	AGG Arg>	GAT Asp>		TTG Leu>		GGA Gly>	910	GAT Asp>	* 096	GGG Gly>	ACT Thr>
	CAA Gln	CGT		AGC Ser	860	TTG Leu	6	GCT Ala		GCT Ala	TCC
	TCT Ser	Sp Sp	810	GAG Glu	ω	TAT Tyr		AGG Arg		GAT Asp	OO ACT Thr
710	TTA Leu	760 AAG G1 Lys A8		ATG Met		GAA Glu		CCA Pro	950	GAA Glu	1000 GCG AC Ala Th
7	GCT	GAT Asp		GTT Val	850	GCA Ala	906.	GAT Asp	0.	CTG	CAT
	AGG Arg	TGG Trp	800	TTG Leu	8	ATT Ile		ACT Thr		AGT Ser	GCT Ala
0	TGC Cys	750 CCG Pro	ω	GTA Val		ATT Ile		ATG Met	940	AGC	990 AAT Asn
700	GCC Ala	AGG Arg		GGA Gly		CCG Pro	890	CAT His	9	GAG Glu	ATA Ile
	GTT Val	TCA Ser	0	GCT	840	GCG Ala	~	TAT Tyr		ATT Ile	TAC Tyr
	TTC Phe	740 GCC	790	$_{\rm GGG}$		GGA Gly		GCT Ala		TGC	980 GTC AAT Val Asn
069	GGA Gly	7 ACT Thr		GAA Glu		CGA	880	GAT Asp	930	TCT	
	GGA Gly	CAG Gln		GGC Gly	830	AAA Lys	8	TGT Cys		TCC	GAG Glu
	TTA Leu	CCT Pro	780	* ATG Met	ω	ATG		AAT Asn		GTC	970 CCT GAA Pro Glu
089	666 Gly	730 GAC CCT ASP Pro		GTG Val		GCA Ala		GTC Val	920	GGT Gly	
o	ATT Ile	GAT Asp		TTT Phe	820	CAT His	870	GCA Ala		CTT Leu	TCA
	CCA Pro	AAT Asn	70	GGT Gly	8	GAA Glu		GGT Gly		666 G1y	GTC Val

1050	G GTT TTC AAG s Val Phe Lys>	1100	G ATC GGA CAC t Ile Gly His>	1150	A ATT AAG GGA r Ile Lys Gly>	1200	C AAT CCC GAG e Asn Pro Glu>	1240 CAG CAA CAT GAA Gln Gln His Glu>	1290	GGC CAC AAC TCA Gly His Asn Ser>	1340	GGT TCA AAT GCA
	AAG AAG Lys Lys	1090	AAG TCG ATG Lys Ser Met	1140	GCG ACA	1190	CAA TTC	AAG Lys		GGA Gly	1330	CTC
1040	AAT GCC ATC Asn Ala Ile	. 10	ACT		GCC ATT Ala Ile	1180	ATA AAC Ile Asn	1230 GCC AAC AAG Ala Asn Lys	1280	GGA TTC Gly Phe	H	A TGA TTA
30	ATA Ile	1080	AAT GCA Asn Ala	1130	CTT GAA	11	CCC AGC	GTT Val	1270	TCA TTC	1320	AAG CCA Lys Pro
1030	GCC GAG Ala Glu		ACA ATC Thr Ile	20	GGG GGT Gly Gly	1170	CTT CAT Leu His	1220 GAC ACA ASP Thr	12	TCA AAT Ser Asn		GCC TTC Ala Phe
1020	GAT CTT Asp Leu	1070	GAA ATC Glu Ile	1120	GCA TCA Ala Ser		GGC TGG Gly Trp	.0 GAA TTC Glu Phe	1260	GCT ATC Ala Ile	1310	TTC TCA Phe Ser
Н	GCT GGG Ala Gly		ACC AAG Thr Lys	1110	CTT GGA Leu Gly	1160	ACC ACC Thr Thr	1210 TCA GTG G2 Ser Val G3	7	AAT GTT Asn Val	00	GTA GCT Val Ala
10	CTT (Leu A	1060	AAC A	H	TGT Cys		ATA Ile	CCA	50	GTG Val	130	GTT Val

FIGURE 2

AATTTGTTGC TGAGACAGTG AGCTTCAACT TGCAGAGCAA TTTTTTACAT GCCTTGTCGT

TATTAGAAAG AACGAGGCAA GATTTTTGTTT CATGTTTGTG TTTGTATTAC TTTCTTTTTG CCCTTGTCAA TGGCATTTAA GATAAGCTTA TAAAAAAAA AAAAAAAA AAAACTCGAG GGGGGGCCCG GTACCCAATT CGCCCTATAG TGAGTCGTAT GACAATTCAC TGTCCGTCGG CGGAAGAGCG TAATACCGGG ATAGTTCCTT GATAGTTCAT TTAGGATGTT TTACTGCAAT AATCGAAGAT TATTTCCATT CTAATCCAGT CTCCGNCGAG TTTGAGAATC TATCTGTTTG

FIGURE 2 5/5

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0 *	ACTAGTGGAT	120	GCTCAGGTGT	ACG TGG Thr Trp		CGT TCC Arg Ser	0	CTC TCC Leu Ser	310	CCT TGC Pro Cys	360	TTC GGA Phe Gly
20	CCGCTCTAGA AC	110	GGTCGGCTCA G	160 TTC TGT Phe Cys	210	GAC CCA (ASp Pro	260	AGG ACT (Arg Thr		CTC GAT	. 0	CTC Leu
40		0		TCC CCT Ser Pro		AAC Asn	0	CGC CGG Arg Arg	300	TGC	350	TTC GCT TCC Phe Ala Ser
ゼ	GCGGTGGCGG	100	TTCTTACTTG	50 GCG Ala	200	TCC Ser	250	CGT		TTC Phe		GGA Gly
30))))	06		ATG GTT Met Val		T TCA r Ser		c TCC u Ser	290	C ACC r Thr	340	T AAC p Asn
` ,	ACAAAAGCTG GAGCTCCACC		GGCACGAGTT	IO TGC Cys	190	CCC ACT Pro Thr	240	CGC CTC Arg Leu		GGA TCC Gly Ser		GGG GAT Gly Asp
20	GCTG GA	80		14 GCT TCT Ala Ser		TGC ATG Cys Met		GG CTC	280	CTC CGC Leu Arg	330	TTC CTC Phe Leu
	ACAAAA		GCAGGAATTC	130 GCG ACC Ala Thr	180	GCA Ala	230	AAG CGG Lys Arg	2	TCC		CGC
10	ACTAAAGGGA	70	торрегост	ATG Met		GTA GCT Val Ala		TCC CAC Ser His	270	CAT TGC His Cys	320	CAG CAA Gln Gln
	ACTA		۵۵۵۵	TCCA	170	CTC	220	CTT	2	TCC		AAC Asn

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ACT Thr		GAA Glu		GTG Val		TAC Tyr	* 009	AAC Asn	TCT Ser	•	GAC
CGC		CAG Gln	200	GTT Val	550	GTT Val	•	GAG Glu	AAG Lys		ATG Met
GGC Gly	450	GCA Ala	20	GTA Val		GAT Asp		ATA Ile	ATC Ile	069	AGG
400 CTC Leu	4	CCT Pro		CGA Arg		CCC Pro	290	GAG	640 GAG Glu		GAG Glu
AGG Arg		CAA Gln		CAA AGG Gln Arg	540	CAT GAC (His Asp	5	AGT Ser	GGA Gly		TCC
CTG	0	ATG Met	490	CAA Gln	.,	CAT His		ATA Ile	GCC Ala	089	TTC
390 CAC His	440	GCT Ala		AAG Lys		GGC Gly		GGC Gly	630 ATT Ile	9	AAG Lys
3 GGC G1y		GTG Val		ACC Thr	530	CTA Leu	580	AGT Ser	AGA Arg		CCA
CGC Arg		GCT	480	GCT Ala	ζζ	CCT Pro		ATA Ile	ACG		GCC Ala
0 AAT Asn	430	ATG Met	Ý	CCT		ACT Thr		GGA G1y	20 CCC Pro	670	GTG Val
38 TCA Ser		GTC Val		AAA Lys		GTG Val	570	GAC	620 TTT CCO Phe Pro		TGG Trp
CGT Arg		GAG Glu	0	AAG Lys	520	GTG Val	2,	CTA	CAG Gln		GGC Gly
CTT Leu	420	GGG Gly	470	AAT Asn		GGC Gly		CTC	TCT Ser	099	5 Q
370 CCT Pro	4	TCC Ser		ACA Thr		ATG Met	260	AAT Asn	610 TGC Cys		ACA Thr
AAG Lys		CAT His		TCC Ser	510	GGT Gly	2	AAC Asn	GAC Asp		TCC
TCC	410	TCC	460	GTC Val	u ,	ACA Thr		TAC	TTC	650	TTT Phe

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FIGI	

GAT Asp		TGT	840 *	GAT	TGT Cys		GAC Asp		ACA Thr	•		GAA Glu
	790	AAG Lys	8	AGC Ser	TTT Phe		ATG Met	086	GCA Ala		1030	GGC Gly
TTA Leu		AGA Arg		TTC Phe	CCC	930	GCA Ala	9	TGT Cys			AAA Lys
GCA Ala		AAA Lys	0	GTA Val	880 AGT Ser		CTT Leu		GCC Ala	•		ATC Ile
AAA Lys	780	AAT Asn	830	AAG Lys	ATC Ile		ATT Ile		ACT Thr		1020	ATA Ile
AAG Lys	7	CTC		ATG Met	AAG Lys	920	TCC GCT Ser Ala	970	TCA		Ţ.	AAC CAC ATA Asn His Ile
GGC		GAG Glu		GGT Gly	870 AAG Lys	92			ATA Ile			AAC
CA La	0	AAA Lys	820	GGC Gly	8 TAT TYY		GGA Gly		TCG		1010	GCG Ala
ACT Thr	770	ATG Met		TTG	TCA		ATG Met	096			10	GCT
CTG Leu		GCG Ala		GGA Gly	860 G ACT g Thr	910	AAT Asn	-	AAC Asn	•		AAT Asn
* ATG Met		GAT Asp	810	TCC	AG		ACA Thr		CCT			CTG
TAC Tyr	160	GAA Glu	ω	GGC Gly	CTG		ACC Thr	950	GGC G1y		1000	ATA Ile
CTT		ACT Thr		ATT Ile	GCT	006	TCT	9	ATG Met			TGT Cys
ATG Met		ATC Ile	0	CTC	850 GAA Glu	01	TTT		${ t TGG}$	•		TTC
TTC Phe	750	GGA Gly	800	GTT Val	ATT Ile		CCT		GGA Gly		066	AAC Asn
AAG Lys	7	GGT Gly		GGA G1y	TCC	068	GTA Val	940	TTG		-	AGT
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FIG

1080	GTT Val	AAT Asn		TTT Phe		CAT His		AGT Ser	1320	GCT Ala	TCG
10	CCT	AAT Asn		GGA Gly	0	GAG Glu	1270	$_{\rm GGG}$	13	GGA Gly	GTC Val
	TTA Leu	AGG Arg	1170	GAT Asp	122	TTA Leu	Н	${ m GGT}$		GAA Glu	GGA G1y
0	GTT Val		11	CGT Arg		GAG Glu		CTA Leu	0]	CCT	1360 ; TCC
1070	GCC Ala	1120 TCA CAG Ser Gln		AAT Asn		GAG Glu	1260	TTT Phe	1310	CAC His	CAG
	GCG Ala	TTG Leu	0.5	AGT Ser	1210	CTT (Leu (12	GAA Glu		CCT Pro	GCT Ala
	GAT Asp	1110 A GCT G Ala	1160	GAC	•	CTT		GCG Ala		GAG Glu	350 TTG Leu
1060	TCG Ser	11 CGA Arg	. •	TGG Trp		TTA Leu	0.0	\mathtt{TAT}	1300	ACC	1350 GCC TTC Ala Leu
7	66C 61y	TGC Cys		CCA Pro	1200	GTT Val	125(ATT Ile		ATG Met	AAG Lys
	GGT Gly	1100 GTA GCA Val Ala	1150	AGA Arg	13	GGA Gly		ACC Thr		CAC	1340 ATA GAG Ile Glu
1050	TGT Cys	1100 GTA GC Val A	(1	TCG Ser		GCT Ala		GCA Ala	1290	TAC	
10	CTT Leu	TTC		GCT Ala	06	GAA GGA Glu Gly	1240	$_{\rm GLY}^{\rm GGT}$	Ä	GCC Ala	TGC
	ATG Met	$_{\rm GGT}$	1140	AAA Lys	119(` '	AGA Arg		GAC Asp	CTC
0	* ATG Met	1090 GGA Gly	Ξ	ACC Thr		GGA Gly		AAA Lys	30	TGC Cys	1330 ATC Ile
1040	GAC Asp	1 TTG Leu		CCT		ATG Met	1230	AAG Lys	1280	ACT Thr	GTG Val
	GCA Ala	GGT G1y	1130	GAC Asp	1180	GTG Val	1;	GCA		TTC Phe	GGT G1y

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TCC ACT CCT GCT Ser Thr Pro Ala	1460	GGC CAA AAC Gly Gln Asn	1510	CAC CTT CTT His Leu Leu	1560	GCA ATA AGG Ala Ile Arg	GAC GAA GGC Asp Glu Gly	1650	CTG AAG GTC Leu Lys Val	1700	AAC TCA TCC Asn Ser Ser
ACT Thr		TGT TTC Cys Phe	1500	ATC GGT Ile Gly	1550	GTT CAG Val Gln	1600 GAA GAC CCG GAC Glu ASP Pro ASP	\leftarrow	GAG AAA Glu Lys		GGC CAT Gly His
CAT GCA His Ala	1450	GCC CAC Ala His	ij	TCG ATG Ser Met		GCA GTA Ala Val	90 TTG Leu	1640	AAG AAG Lys Lys	1690	TTC GGC
AAT GCG Asn Ala	1440	A GCT CTC	1490	C ACC AAA	1540	A GCA GTT u Ala Val	ATT Ile		c GGC CCT 1 Gly Pro	1680	A TTT GGG r Phe Gly
r TAC ATA n Tyr Ile		A TAC CAA u Tyr Gln		G AAT TCC 1 Asn Ser	1530	C GTA GAA y Val Glu	1580 T CCA AAT s Pro Asn	1630	G CTC GTC		C AAT TCA er Asn Ser
* C GTA AAT p Val Asn	1430	C AAG GAA e Lys Glu	1480	G AGA GTG u Arg Val		T GGT GGC a Gly Gly	570 TGG ATC CAT Trp Ile His	1620	GCA AAA CTG Ala Lys Leu	1670	GGT TTG TCC Gly Leu Ser
s GAA GAC y Glu Asp		A GAT ATC Y ASP Ile	1470	r GAG CTG r Glu Leu	1520	A GGA GCT y Gly Ala	1 GGA G1Y		GAT Asp	0	GTC Val
AGG Arg	1420	GGA Gly	, ,	AGT		GGA Gly	ACA	1610	GTG Val	1660	AAG Lys

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00 /	rtga	1820	CGA	1880	TTT	1940	AAT	2000	GAA		
	GAGTCTTTGA	⊣	GGCTACT	T	TTGTCCC	7	CTTTTCG	2	ATATTTT		
	TGGAAGCCGA	1810	GAGATAGACC	1870	AGATCACTGC	1930	TATTTCGAG	1990	TTTGTAATGC		AAAAA
K	AAC TAG A AAAGAGTCTG Asn ***	1800	CTTCTTATGC CTCTGAAACC GAGATAGACC GGCTACTCGA	1860	TGGTGTTAAG	1920	GAGGTAGTCG	1980	TCTAAGATCA	2040	AAAAAAAA
	C AAC TAG A s Asn ***	1790	CTTCTTATGC	1850	TTGCCGGTAT	1910	AGCTTTAACC	1970	ATGTGTTTCT	2030	AAATAAAAA
	GCC CCC TGC Ala Pro Cys	1780	GAACTCATGC ACGTTAGTAG	1840	GGGGATGCCA AAGATACTCC TTGCCGGTAT TGGTGTTAAG AGATCACTGC TTGTCCCTTT	1900	TATITICITC ITCITITGAG AGCITTAACC GAGGTAGICG TATITICGAG CITITICGAAT	1960	TATCGGATCA ATGTGTTTCT TCTAAGATCA TTTGTAATGC ATATTTTGAA	2020	TCAGTATGCA AAATAAAAA AAAAAAAAA AAAAAA
	ATA CTA TTT Ile Leu Phe	1770	GAACTCATGC	1830	GGGGATGCCA	1890	TATTTTTT	1950	ACATGTTCGT	2010	AAACCACATC

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	09	CTACACCTCC	120	GCTCAATCGA	180	AGTTACCACA	ATG Met>		AAT Asn>		TGT Cys>	370	ACA Thr>
.*		TAC		3CTC		AGTT.	GGA Gly		AAT Asn	320	GAT Asp	n	TCC
	20		110		170	GA i	220 'G ACT	270	TAC		TTT Phe		TTC
		TGAC	←-1	9922	1	ACAG	GT Va	•	TTC Phe		ACC Thr		Ser
		GCCATGACTA		GGCACCGGAG		CTGCACAGGA	GTT Val		GTT Val	310	GAG	360	AAG Lys
	40		100	GCA	160		3TA Val	260	GAT	31	ATA Ile		ÀTC Ile
		TTCGAGCCCT		ACCC		GCTCTGCAAC	210 CGA Arg		CCT Pro		GAG Glu		GAG Glu
				ACC	.•		CGG Arg		GAC Asp	,	AGT Ser	350	GGA Gly
	30	CCTCGCCTGC	90	ລອລລ	150	AATGGCTGTG		0	CAT His	300	ATA Ile	()	AGA ATT GCT GGA Arg Ile Ala Gly
		rcgc		CCAT		TGGC	200 ATC AAA CAG Ile Lys Gln	250	GGC Gly		GGC Gly		ATT Ile
	C	CC.	0	G GC	0	C AA	ATC Ile		CTA Leu		AGT	0	
	20	rctt.	80	TCCA	140	GAGG			CCT Pro	290	ACG Thr	340	CCT.ACG Pro Thr
		CACC		CGGA		CCGGGGGAGGC	0 CCA Pro	240		. (2)	GGA Gly		CCT
1921	10	3G .T(7.0	GT T	130		190 AAG CCA AGT Lys Pro Ser		GTG Val		GAT Asp		$ ext{TTT}$
L to		ACGA(•	CCTT	-	ממממ	AAG Lys		GTG Val	0	CTT Leu	330	CAA Gln
је: 1		CGGCACGAGG TCACCTCTTA		SCATCCTTGT TCGGATCCAG GCCCATCCGC ACCACCCGCA		GCTTCCCCTT	AAG Lys	30	GGT	280	CTG		GCT Ala
Rang				0		J	/ 7 . 7	2					
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IGURE 4

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	420	ATG Met>	ATC Ile>		CTC Leu>		GAA Glu>	610	TTC Phe>	* 099	TGG Trp>	TTT Phe>
1		TTC	GGA		GTT Val	260	ATT Ile	9	CCT Pro		GGA Gly	AAC Asn
		AAG	$^{ m GT}_{ m J}$	510	GGA Gly	ഥ	GCC Ala		GTA Val		TTG Leu	700 ACG AGT Thr Ser
	410	GAC	460 AAT G Asn G		TGC Cys		GAT Asp		TGT Cys	029	GAC Asp	
	4	ATG (ACA		AAA Lys	0	AAT Asn	009	TTT Phe	Ψ	ATG Met	GCA Ala
		AGG A	TTA	200	AGA Arg	55	TTC Phe		CCC Pro		GCA Ala	TGT Cys
	0	AAG I	450 GCA A	Ŋ	AAA AGA Lys Arg		GTA Val		AAT Asn	0	CTT	690 GCT Ala
	400	TCC /	AAA Lys		GAT		AAG Lys	290	ATG Met	640	ATG	ACT Thr
		CTC	AAG . Lys	0	CTA	540	ATG Met	Ŋ	AAG Lys		GCT	TCT Ser
		AAG (Lys	440 GGC Gly	490	GAG Glu		GGA Gly		AAG Lys		TCA	680 ATA Ile
	390	CCG Pro	GCC Ala		AAA Lys		GGT Gly	0	\mathtt{TAT}	630	GGA Gly	TCG Ser
	•	GCC	ACT		ATG Met	530	ATG Met	580	TCA		ATG Met	TAC
		GTG Val	rG eu	480	grg Val	2	GCA Ala		ATT Ile		AAT Asn	670 CCC AAC
	380	TGG (Trp	430 ATG C		GAT		TCA Ser		AGG Arg	620	ACA Thr	67 CCC Pro
	ñ	GGT '	TAC		GAA Glu	0	GGC Gly	570	CTA	v	ACC Thr	GGC G1v
		GAT (Asp (CTT Leu	470	ACC Thr	520	ATT		GCC Ala		GCT	ATG

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	GTG Val>		GGA G1y>	20	ACT Thr>	*	GGG G1y>	AAA Lys>		TGC Cys>	•	ATT Ile>	
	GAT	800	ATG Met	85	CCT		ATG Met	AAG Lys		ACT Thr	1040	GTG Val	
750	GCA A	∞	GGT Gly		GAC Asp		GTT Val	10 GCA Ala	066	TTC Phe	1(GGA	
	GAA (ATT		GCC	890	TTT Phe	940 CAT GC His A.		AGT Ser		GCT	
	GGC (Gly (0.	CCT. Pro.	840	AAT Asn	∞	GGA Gly	GAG Glu		GGA Gly	30	GGA Gly	
740	AGA (Arg (790	ATA Ile		AGA Arg		GAT Asp	TTA Leu	086	GGT Gly	1030	GAT	
7,	ATC I		ATC		CAG Gln	0	CGT Arg	930 GAG Glu	, 01	CTA		CCT	
	ATA . Ile		GTA	830	TCA	880	AAT Asn	GAG Glu		TTT Phe		CAC	
	CAC	780	GCG	∞	TTG		AGT Ser	CTA	070	GAA Glu	1020	CCT	
730	AAC (Asn		GAT		GCT		GAC Asp	920 CTA Leu	6	GCA		GAG Glu	
	GCG		TCA	0	CGA Arg	870	TGG Trp	9 CTA Leu		TAC Tyr		ACC Thr	
	GCT	770	GGC Gly	820	TGC		CCA	GTG Val		ATT Ile	1010	CAC ATG His Met	
720	AAT Asn	7	GGG		GCA Ala		AGA Arg	10 GGA G1y	096	ACT Thr	1(
•	CTG	, .	TGC		GTT Val	098	TCA Ser	910 GCT G		GCG Ala	٠.	TAC	
	ATC Ile	0	CTT Leu	810	TTT Phe	∞	GCT Ala	GGA G1y		GGT Gly	00	GCC	
710	TGT 7	760	ATG (GGT Gly		AAA Lys	GAA Glu	950	AGA Arg	1000	GAT Asp	

FIGURE 4

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AAA GTG AAT TCT ACC AAA TCA ATG ATT GGT CAC CTT CTC GGA GCA GCC Lys Val Asn Ser Thr Lys Ser Met Ile Gly His Leu Leu Gly Ala Ala> AAA GAG TAC CAA GCT CTT ATC CAC TGT TTC GGC CAA AAC AAC GAG TTA Lys Glu Tyr Gln Ala Leu Ile His Cys Phe Gly Gln Asn Asn Glu Leu> GGT GGT GTG GAA GCA GTT TCA GTA GTT CAG GCA ATA AGG ACT GGG TGG Gly Gly Val Glu Ala Val Ser Val Val Gln Ala Ile Arg Thr Gly Trp> Ile His Pro Asn Ile Asn Leu Glu Asn Pro Asp Glu Gly Val Asp Thr> AAA TTG CTC GTG GGC CCT AAG AAG GAG AGA CTG AAC ATT AAG GTC GGT Lys Leu Leu Val Gly Pro Lys Lys Glu Arg Leu Asn Ile Lys Val Gly> Thr Ser Thr Pro Ala Gly Asp Ile> Glu Asp> 1140 ATC CAT CCG AAT ATT AAT TTG GAA AAC CCA GAT GAA GGC GTG GAT ACC GAA GAC GTA AAT TAC ATA AAT GCA CAT GCC ACA TCC ACT CCA GCT GGA GAT ATC Val Asn Tyr Ile Asn Ala His Ala Thr Ser Thr Pro Ala Gly Asp Iles 1280 CTC TGC ATA GAG AAG GCT TTG GCT CAG TCA GGA GTC TCT AGG Leu Cys Ile Glu Lys Ala Leu Ala Gln Ser Gly Val Ser Arg 1370 1130 1320 1120 1310 1260 1210 1110 1350 1300 1250 1200 1240 1190

FIGURE 4

ATA CTC TTC Ile Leu Phe>	1480	TCAAA	1540	CATGCCCATG	1600	GGCGACACAG	1660	TTTCTGAAAT	1720	AGTCAGTGAA GAAGAGAACA	1780	TTTATCGCCG	1840	TTTTCTCTTG ATCATTGGAG
1420 TCG TCC Ser Ser	1470	ATTCTACTCA ATCTATCAAA	1530	TAGCTCCTTA CGTCTCTAGA CATGCCCATG	1590	GAGTACTCAT	1650	CTATTCATTA TCCCATTTTT TTTCTGAAAT	1710		1770	TGCTCTCTAT	1830	
1410 GGG CAC AAC Gly His Asn	1460	CATGTGGGA ATTCT	1520	TAGCTCCTTA	1580	ATGACGGATT	1640	CTATTCATTA	1700	CGTTTCATCG	1760	CCCTTTGTTT	1820	GACTGGTTTG
1400 GGG TTT GGT G Gly Phe Gly G	1450		1510		1570	AGTCGGAACC	1630	TGTTAGAGCA	1690	CTCCCTCCTT ACGGTAGTTG TACTTTCGAG	1750	TAACCATTTG	1810	TTTTGTGGGT TAAAATTTGT AAAACTAGAC GACTGGTTTG
TCA TTC Ser Phe	1440	TAC AAC TAG GGCGTTT Tyr Asn ***>	1500	TGAGGACTCC AGCATGTTGG	1560	CGGGAGCTGT	1620	TTGCTAGAAT	1680	ACGGTAGTTG	1740	GGGCACGTAG	1800	TAAAATTTGT
1390 TTG TCT AAT Leu Ser Asn	1430	GCC CCT TAC Ala Pro Tyr	1490	GCTGAAGTTT	1550	AGTTTTGTGT	1610	GATATACTCC	1670	CTCCCTCCTT	1730	AAGCTAACTC	1790	TTTTGTGGGT

FIGURE 4 5/6

FIGURE 4 6/6

ATGTATGGCC ATATTTGCCT TTCATTGATG ATAAAAAAA AAAAAAAA AAAAAAAAA 1890 1880 1870 1860

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09	120	169	217	265	313	361	409	457	505
CIGGIACGCC IGCAGGIACC GGICCGGAAI ICCCGGGICG ACCCACGCGI CCGICTICCC	ACTCCGATCG TTCTTCTTCC ACCGCATCTC TTCTCTTCTC	CGCCGCC ATG CAT TCC CTC CAG TCA CCC TCC CTT CGG GCC TCC CCG CTC Met His Ser Leu Gln Ser Pro Ser Leu Arg Ala Ser Pro Leu 1	GAC CCC TTC CGC CCC AAA TCA TCC ACC GTC CGC CCC CTC CAC CGA GCA Asp Pro Phe Arg Pro Lys Ser Ser Thr Val Arg Pro Leu His Arg Ala 15	TCA ATT CCC AAC GTC CGG GCC GCT TCC CCC ACC GTC TCC GCT CCC AAG Ser Ile Pro Asn Val Arg Ala Ala Ser Pro Thr Val Ser Ala Pro Lys 35	CGC GAG ACC GAC CCC AAG AAG CGC GTC GTG ATC ACC GGA ATG GGC CTT Arg Glu Thr Asp Pro Lys Lys Arg Val Val Ile Thr Gly Met Gly Leu 50	GTC TCC GTT TTC GGC TCC GAC GTC GAT GCG TAC TAC GAC AAG CTC CTG Val Ser Val Phe Gly Ser Asp Val Asp Ala Tyr Tyr Asp Lys Leu Leu 65	TCA GGC GAG AGC GGG ATC GGC CCA ATC GAC CGC TTC GAC GCC TCC AAG Ser Gly Glu Ser Gly Ile Gly Pro Ile Asp Arg Phe Asp Ala Ser Lys 80	TTC CCC ACC AGG TTC GGC GGC CAG ATT CGT GGC TTC AAC TCC ATG GGA Phe Pro Thr Arg Phe Gly Gly Gln Ile Arg Gly Phe Asn Ser Met Gly 95	TAC ATT GAC GGC AAA AAC GAC AGG CGG CTT GAT GAT TGC CTT CGC TAC Tyr lle Asp Gly Lys Asn Asp Arg Arg Leu Asp Asp Cys Leu Arg Tyr 120

553	601	649	697	745	793	841	888
GCC Ala	666 Gly	CTT Leu	GCC Ala 190	ATG Met	TGC Cys	ATG Met	GGC Gly
GGT Gly	GTT Val	TCT Ser	TAT Tyr	CTG Leu 205	TAC	CTT Leu	GGA Gly
CTC Leu 140	CTG	CAA Gln	CCC Pro	$_{\rm G1Y}$	AAC Asn 220	GAT Asp	TTG
GAT Asp	GTG Val 155	GTT Val	ATC Ile	CTC	TCC	GCT Ala 235	666 G1y
GCC Ala	GGA Gly	GGG Gly 170	TTC Phe	GAA Glu	ACT Thr	GAG Glu	ATT Ile 250
GAC Asp	GCC Ala	GAC Asp	TTC Phe 185	ATT Ile	GCC Ala	$_{\rm G1y}$	CCA Pro
GAG Glu	AGA Arg	TCT	CCT Pro	GCT Ala 200	TGT Cys	CGT Arg	ATT Ile
CTT Leu 135	GAG Glu	TTC Phe	ACC Thr	CTC	GCA Ala 215	CGC Arg	ATC Ile
TCT	AAG Lys 150	GTC Val	ATC Ile	CTG	ACT Thr	ATC 11e 230	GCA Ala
AAG Lys	GAC Asp	ACT Thr 165	AAA Lys	GCC Ala	TCC Ser	CAT His	GCC Ala 245
AAG Lys	ATC Ile	CTG	CGG Arg 180	TCT Ser	ATT Ile	AAT Asn	GAG Glu
GGG G1γ	AAG Lys	GGT Gly	CAC His	GGG G1Y 195	TCA Ser	GCT Ala	ACT Thr
GCC Ala 130	TCC Ser	GGT Gly	GGT Gly	ATG Met	TAT Tyr 210	GCT Ala	GGC
GTC Val	CTC Leu 145	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT Ala 225	GGA Gly
ATT Ile	CGC Arg	GGA G1y 160	GAG Glu	ACA Thr	CCA Pro	CAT His	GCT Ala 240
TGC Cys	GAC	ACA Thr	ATC Ile 175	ATT Ile	GGC Gly	TTC Phe	ATT Ile

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937	985	1033	1081	1129	1176	1224	1272
T H O	د ۲	CGA Arg	GAT Asp	TCT Ser	GTC Val 350	GCC Ala	AAA Lys ··
ACT Thr 270	GAA Glu						
CAG Gln	GGT G1y 285	AAA Lys	TGT Cys	TCC	GAG Glu	CTC Leu 365	ATC Ile
CCT	ATG Met	ATG Met 300	AAC Asn	GTC Val	GAA Glu	GAT Asp	GAT Asp 380
GAC Asp	GTG Val	GCA Ala	ATC Ile 315	GGT.	CCT Pro	GGG Gly	AAG Lys
GAT Asp	TTT	CAT His	GCA Ala	CTC Leu 330	TCA Ser	GCT Ala	ACA Thr
AAC Asn 265	GGT Gly	GAA Glu	GGT Gly	$_{\rm GLY}^{\rm GGT}$	GTC Val 345	CTA Leu	AAC
AGG	GAT Asp 280	TTG Leu	GGA Gly	GAT Asp	66C G1y	ACT Thr 360	AAG Lys
CAA Gln	CGT Arg	AGC Ser 295	TTG Leu	GCT Ala	GCT Ala	TCT Ser	TTC Phe 375
TCT Ser	GAC Asp	GAG Glu	TAT Tyr 310	AGG Arg	GAT Asp	ACT Thr	GTT Val
CTG Leu	AAA Lys	CTG Leu	GAG Glu	CCA Pro 325	GAA Glu	GCG Ala	AAG Lys
GCT Ala 260	GAT Asp	GTG Val	GCA Ala	GAC Asp	CTT Leu 340	CAT His	AAG Lys
AGG Arg	TGG Trp 275	TTG Leu	ATT Ile	ACT Thr	AGC Ser	GCT Ala 355	ATC Ile
TGC Cys	CCC	GTG Val 290	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala 370
GCT Ala	AGG Arg	GGA Gly	CCT Pro 305	CAC His	GAG Glu	ATA Ile	AAT
GTG Val	TCT Ser	GCT Ala	GCA Ala	TAT Tyr 320	ATT Ile	TAC Tyr	ATA Ile
TTT Phe 255	GCC Ala	GGT Gly	GGA Gly	GCT Ala	TGC Cys 335	AAT Asn	GAG Glu

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sa GCC TCT GGA 1320 ly Ala Ser Gly 95	CC GGC TGG CTT 1368 hr Gly Trp Leu	GAG TTC	3 1)	GCG ATC Ala Ile 445	GCG ATC TCG Ala Ile Ser 445 TTC TCG GCT Phe Ser Ala	ATC TCG Ile Ser 445 TCG GCT Ser Ala	ATC TCG Ile Ser 445 TCG GCT Ser Ala	ATC TCG Ile Ser 445 TCG GCT Ser Ala CGTTTCCCTTC
AC TGT CTT GGA is Cys Leu Gly 395	sa ara aac acc ly ile Asn Thr 410	GAG CCA TCC GTG Glu Pro Ser Val	4 1	GAA GTT AAT GTT Glu Val Asn Val 440	GTT AAT Val Asn GTC GTG Val Val	AA GTT AAT GT 1u Val Asn Va 40 CA GTC GTG GC er Val Val Al CTTGTC ATTGAG	AC GAA GTT AAT GTT GCG is Glu Val Asn Val Ala 440 AC TCA GTC GTG GCT TTC sn Ser Val Val Ala Phe 55 GGCACTTGTC ATTGAGAGTA AAAAAAAGTA AGGATTATCA	AA GTT AAT GT' 1u Val Asn Va 40 CA GTC GTG GC' er Val Val Al. CTTGTC ATTGAG. TTATTT TAAAAA
ATG ATC GGA CAC Met Ile Gly His 390	ACT ATT AAG GGA Thr Ile Lys Gly 405	TTC AAT CCT Phe Asn Pro		CAG CAA CAC Gln Gln His	CAG CAA CAC Gln Gln His GGC CAC AAC Gly His Asn	CAG CAA CAC Gln Gln His GGC CAC AAC Gly His Asn 455 ATTTCACAA GGC	G CAG CAA CAC GAS GIN GIN His GI 44 A GGC CAC AAC TC Y GIY His Asn Se CATTTCACAA GGCAC	CAG CAA CAC GA Gln Gln His Gl GGC CAC AAC TC Gly His Asn Se STTCTATGTA AAAA/ SAATGAAATT ATAT
ACT AAG TCA Thr Lys Ser	GCT ATA GCG Ala Ile Ala	ATT AAT CAA Ile Asn Gln 420	j	AAC AAG Asn Lys 435	AAC AAG Asn Lys 435 GGA TTC Gly Phe 450		AAC AAG AAG ASD Lys Lys 435 GGA TTC GGA Gly Phe Gly 450 TGA TTACC C.	AAC AAG AAG ASD Lys Lys 435 GGA TTC GGA Gly Phe Gly 450 TGA TTACC C. TGA TTACC C.
ATT AAT GCA Ile Asn Ala 385	GGT CTT GAA Gly Leu Glu 400	CAT CCC AGC His Pro Ser 415		ACT GTT GCC Thr Val Ala	GTT Val TCA Ser	GTT Val TCA Ser AAG Lys	GTT (Val ANG AAG LYS	ACT GTT GCC Thr Val Ala AAT TCA TTT Asn Ser Phe TTC AAG CCA Phe Lys Pro TCAAACCCAT

Sequence Range: 1 to 1802

		•									
*	TTATCTCCGC	110 TCC CCT TCC Ser Pro Ser	160	TCC TCC	210	: ATC CGT		3 AAG CGG 5 Lys Arg		GAC GTC Asp Val	350 ATC AGC CTA Ile Ser Leu
20		CTC CAC Leu His		r TCC CCC 1 Ser Pro	200	ccc GTC n Pro Val	250	C CCC AAG p Pro Lys	300	c GGC TCC e Gly Ser	GGC Gly
40	CTTTCCGACC ACATTTCATT TCTTGCCTCG	100 CAA TCC Gln Ser	150	c CTC AAT g Leu Asn		C AGC CTC a Ser Leu		G TCC GAC u Ser Asp	290	C GTC TTC r Val Phe	340 C GAG AGC Y Glu Ser
30	C ACATT	90 GC C ATG Met	140	C TTC CGC Phe Arg	190	r CGC GCC g Arg Ala	240	G CGC GAG s Arg Glu		C GTC TCC u Val Ser	C TCC GGC u Ser Gly
8	TTTCCGAC	ວອວວອວວອວວ 06		GAG CCC		CTC CGT	230	CCC AAG Pro Lys	280	GGC CTC	330 3 CTG CTC 5 Leu Leu
20	CGCGTCCGGG C		130	CCT CTC Pro Leu	180	CGC CCC		TCC GCC		GGC ATG	320 TAC GAC AAG Tyr ASP Lys
10		70 CGCTCCTCCG CCGTCGTTCG		CCC TCC Pro Ser	170	GCT CTC Ala Leu	220	ACC GCC Thr Ala	270	ATC ACC Ile Thr	TAC
•	GGTCGACCCA	CGCTCCT	120	CTC CGC Leu Arg	7	GCC GCC Ala Ala		GCT GCC Ala Ala	260	GTC GTC Val Val	310 GAC GCC Asp Ala

FIGURE 6 1/5

-	c cAG y Gln	450	c cgg p Arg		G GCT s Ala		T AAG p Lys	T GTC r Val	0	G ATC	069	GCG CTG Ala Leu	
	GGC Gly		GAC Asp		3 AAG s Lys		r GAT e Asp	590 CTA ACT Leu Thr	640	G AAG g Lys			
	GCC Ala		AAC Asn		AAG Lys	540	ATT Ile	CTI		CGG Arg		TCT Ser	
	TTC Phe	o _j	AAG Lys	490	GGC Gly		AAG	GGC Gly		CAC His	089	GGG G1y	
390	AGG Arg	440	GGC Gly		GCC		TCC	GGT Gly	630	GGT Gly	9	ATG Met	
m	ACC Thr		GAC Asp		GTC Val	530	CTC	580 ATG Met		AAA Lys		AAC Asn	
	CCC Pro		ATC Ile	480	ATT Ile	Ω	TCC	GGT Gly		GAG Glu		ACA Thr	9
0	TTC Phe	430	TAC	7.	TGC ATT GTC (Cys Ile Val		CAA Gln	570 GTT GGA ACC Val Gly Thr	620	ATC Ile	670	ATT Ile	FIGURE
380	AAA Lys		GGC Gly		TAC Tyr		GCC GGC Ala Gly	570 GGA Gly	9	CTC		GCC	FI
	TCC		ACG Thr	470	CGC Arg	520	GCC Ala	GTT Val		AAT Asn		TAT Tyr	
	GCT Ala	420	AAC GCG ACG Asn Ala Thr	4	CTC		CTC	CTA Leu		CAG Gln	* 099	ATT CCA Ile Pro	
370	GAC	7			TGC		GAT Asp	560 GGA GTG Gly Val	610	GTT Val		ATT Ile	
	TTC Phe		TTC Phe		GAT Asp	510	GCC Ala			GGG G1y		TTC	
	CGC Arg	0	GGC Gly	460	GAC Asp	u,	GAC	GCC		GAC Asp	650	TTT Phe	
360	GAC Asp	410	CGT		CTC		GAA Glu	AGG	009	TTC TCT Phe Ser	9	CCG	
m	ATC Ile		ATC Ile		CGG Arg	200	CTC	550 GAG Glu	_	TTC Phe		TCC	

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	ACT Thr		ATC Ile	GCG		TCT Ser	930	GAC Asp		GAG Glu	•	TAT Tyr
	TCA		CAT	30 GCT Ala	880	TTA Leu	o)	AAG Lys		ATG Met		GAA Glu
	ATT Ile	780	AAT Asn	830 GAG G(Glu A		GCT		GAT Asp		GTT Val	1020	ATT GCA Ile Ala
730	TCG Ser	7	GCC Ala	ACT		AGG Arg	920	TGG Trp	970	TTG	T	
	TAT Tyr		GCC Ala	GGA Gly	870	TGC	92	CCG Pro		GTA Val		ATT Ile
	AAC Asn	0	GCT	820 GGA G1Y	ω	GCC Ala		AGG		GGA Gly	10	CCG
720	CCA Pro	770	TAT Tyr	GCT		GTT Val		TCA	096	GCT Ala	1010	GCG
7	GGC Gly		TTT Phe	ATT Ile	098	TTC Phe	910	GCC	01	$_{\rm GGG}$		GGA Gly
	ATG Met		TGC	810 CTG ATG Leu Met	86	GGA Gly		ACT Thr		GAA Glu		CGG Arg
0	CTG Leu	760	TAC Tyr	CTG Leu		GGA Gly		CAG	950	$_{\rm G1Y}$	1000	AAA Lys
710	$_{\rm GGT}$		AAC Asn	GAC Asp		TTA Leu	006	CCT	9	ATG Met		ATG Met
	TTG		TCC	O GCT Ala	850	GGT Gly	01	GAT Asp		GTG Val		GCA
	GAT Asp	750	ACT Thr	800 GAG GG		ATT Ile		GAT Asp		TTT Phe	066	CAT
700	ATC Ile	7	GCT Ala	GGT Gly		CCA Pro	068	AAT Asn	940	GGC Gly		GAG Glu
	GCC Ala		TGT	CGA Arg	840	ATT Ile	8	AGG Arg		GAT Asp		TTG
	CTT Leu	740	GCA Ala	790 CGC Arg	۵	GTC Val		CAA		CGT	980	AGC

FIGURE 6 3/5

1030												
1080 1080 1080 1080 1080 1080 1080 1080	AGG Arg		GAT Asp	70	ACT Thr		GTT Val			ATT Ile		
1080 1080 1080 1080 1080 1080 1080 1080		120		11	GCG Ala				ATG Met	LO ACC Thr	1360	TTT Phe
1080 1080 1080 1080 1080 1080 1080 1080	107 GAT ASP	⊣	CTC		CAT His		AAG Lys	*	TCA		` '	
1040			AGT Ser	0.0	GCT Ala	1210	ATT Ile	13				
1040	ATG Met	10		116	AAT Asn	()			ACT Thr		350	ATT Ile
1040 1040 1050 1080 1080 1080 1080 1080 1080 108	060 CAT His	11	GAG Glu		ATA Ile		AAT Asn	20	GCA Ala	1300 GAA Glu	- i	AGC
1040 1040 1050 1080 1080 1080 1080 1080 1080 108	1 TAT TYY		ATT Ile		TAC Tyr	003	ATA Ile	12	AAT Asn			CCC
GGGG GGT GCA GTC AAC TGT GAT U G1y G1y Ala Val Asn Cys Asp 1080 1080 1090 11080 11130 11130 11180 11180 11190 Tr GAT GGG CTT GGT GTC TCC TCG CGG GTC TCA CCT GAA GAG GTC TACT TCA CCT GAA GAG GTC TACT CTT GCT GGG GAT CTT GCC ET Thr Leu Ala G1y Asp Leu Ala 1230 1280 1280 1280 1320 1320 1330 1330 1330 1340 1350	GCT Ala	0	TGC	150		13	GAG Glu		ATC Ile		40	CAT His
G GGA GGT GCA GTC AAC u Gly Gly Ala Val Asn 1080 T GAT GGG CTT GGT GTC a Asp Gly Leu Gly Val 1130 1180 T ACT CTT GCT GAA T ACT CTT GCT GAA TC AAG AAC ACC AAG GAA he Lys Asn Thr Lys Glu 70 GA CAC TGT CTT GGA GCA 1320 AG GGA ATA ACC ACC GGC AG GGA TTA Thr Gly Ala 1320 1320 AG GGA ATA ACC ACC GGC AG GAT Thr Leu Gly Ala 1320 AG GGA ATA ACC ACC GGC AG GGA ATA ACC GGC AG GGA ATA ACC ACC GGC		110	TCG Ser	,1	GTC Val		GCC Ala		AAA Lys	290 GGA Gly	13	CTT
G GGA GGT GCA GTC AAC u Gly Gly Ala Val Asn 1080 T GAT GGG CTT GGT GTC a Asp Gly Leu Gly Val 1130 1180 T ACT CTT GCT GAA T ACT CTT GCT GAA TC AAG AAC ACC AAG GAA he Lys Asn Thr Lys Glu 70 GA CAC TGT CTT GGA GCA 1320 AG GGA ATA ACC ACC GGC AG GGA TTA Thr Gly Ala 1320 1320 AG GGA ATA ACC ACC GGC AG GAT Thr Leu Gly Ala 1320 AG GGA ATA ACC ACC GGC AG GGA ATA ACC GGC AG GGA ATA ACC ACC GGC						90	CTT Leu	1240	ATC Ile	TC		
CG GGA GGT GCA GTC CU Gly Gly Ala Val 1080 TO GAT GGG CTT GGT A ASP Gly Leu Gly 1130 TO GGG GTC TCA CCT 1180 TO AGG GTC TCA CCT 1180 TO AGG AAC ACG AAG TO AAG AAC ACC ACC TO AAG AAC ACC AAG TO AAG AAC ACC AAG TO AAG AAC ACC AACC TO AAG AAC ACC AAG TO AAG AAC ACC AACC TO AACC AAC	AAC Asn		GTC Val	40		119			GAA Glu			
TG GGA GGT GCA TO GGA GGT GCA TO 80 TGAT GGG CTT TASP GIY Leu TG GG GTC TCA TACT CTT GCT TACT CTT GCT TACT CTT GCT TACT CTT GCT TC AAG AAC ACC TC AAG AAC ACC TC AAG AAC ACC TC AAG AAC ACC THIS CYS Leu TA CT CTT TO TACT TO TO TACT TO TACT	0 GTC Val	060	GGT Gly	Η	CCT Pro				AAG Lys	80 GGA Gly	1330	
TG GGA GGG GG	104 GCA Ala	Т	CTT Leu		TCA			230	ACC Thr	0 11		
TG GGA (1080) TG GGA (1080) TG GGG (10 GG) TC AAG (10 His In H	GGT Gly		$_{\rm GGG}$	0.	GTC Val	1180		ij				
Signal of the control	GGA Gly	980	gaT Asp	113	GGG	•			AAG Lys		320	
		10	GCT		GCC Ala		TCT Ser	1220	TTC	1270 GGA Gly	H	AAG Lys

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1410	AAG CAG CAA Lys Gln Gln		GGG CAC Gly His	1510	ATTCT ACTTGGTTCA	1570	TAAATGCCTT	1630	AGCCATTTAG	1690	CTCTGATTTA	1750	GTTATTTAAG		CT
1400	AAC AAA AAG Asn Lys Lys	1450	GGA TTT GGA Gly Phe Gly	1500	TGA ATTCT A	1560	AGCAATTTTT	1620	GTCCTTTGAT AGTTCCTCGA	1680	TAAATCTAGT	1740	TGTTGTCAAT	1800	ATCCAGCTTA
06	ACT GTT GCC Thr Val Ala	1440	AAT TCT TTT Asn Ser Phe	1490	TTC AAG CCA Phe Lys Pro	1550	CAACTTGCAG	1610	GTCCTTTGAT	1670	ATTCCCATTT TAAATCTAGT	1730	GTCATGTTTG	1790	GCTCTAGAGG ATCCAGCTTA
1390	TTC AAC Phe Asn	1430	ATC TCG Ile Ser	1480	TCA GCT Ser Ala	1540	GATAGGGCTT	1600	GAATAGGTCG	1660	ATCGAAGATG	1720	AAGATTTTGT	1780	AAGGGCGGCC
1380	TCG GTG GAC Ser Val Asp		AAC GTC GCT Asn Val Ala	1470	GTG GCA TTC Val Ala Phe	1530	CAGTTGCTGA	1590	CGTAATACCG	1650	таствтаата	1710	AGACCAATGA	1770	ataaagcaaa aaaaaaaaa aaggggggg
1370	CCC GAG CCA Pro Glu Pro	1420	CAT GAA GTG His Glu Val	460 1	AAC TCG GTT Asn Ser Val	1520	AAATGCACAC	1580	GTCGGAAGAG	1640	GATGATGTTT	1700	TGTATTAGAA	1760	ATAAAGCAAA

180 120 AGAGAGAGGG ATCCATCGAA TGCGGCCACC CTCCTTTCAT CTTCGATTCA TTACCATACC GTACGCCTGC AGGTACCGGT CCGGAATTCC CGGGTCGACC CACGCGTCCG CATAAAAAGAG ATTCCGCTGA TCCATTTTCC GCCTTTTCG GGTCTTTCAT CCCAAAGGGT ATCCTTTTCT Pro> Gln Cys Ala Pro Leu> CCT CTC TGT ACG TGG CTC CTT GCC GCC TGC ATG TCT Pro Leu Cys Thr Trp Leu Leu Ala Ala Cys Met Ser> Met Pro Ala Ala Ser Ser> 330 TCC GAC CCT CTT CCG CCT TCC ATC TCC TCT CCT CCA CTA 190 200 210 220 230 ATCCTATCTT CTCAAAGGGT CAGTCAGTTC CCTCCA ATG CCT GCC GCC TCT TCC Ser CGC CGC CGG ATT CTC TCC CAA TGC GCC Arg Arg Arg Ile Leu Ser Gln Cys Ala Ser Phe His Pro Ser Asp Pro Leu Pro Pro Ser Ile Ser 170 370 110 50 320 270 160 100 40 360 310 260 150 30 90 350 300 250 140 20 80 CCC 1 CC TCC TTC CAC TCCLeu Leu Ala Ser CGC CTC 340 Sequence Range: 1 to 2369 CTC GCT 290 130 CGA (၁၅၁ ACC Thr CIG

FIGURE

Arg Arg

Arg Arg Leu Ser

Arg

	٨	^		^		. ^		ري در دي در		٦ \	& &
	GTC Val>	TCC Ser>	520	CGG Arg>	57.0	CTG Len>		CAG Gln>		CAT His>	: ATA / Ile>
	CTC	470 ACA Thr	. 23	CAC His		GCT Ala		AAA Lys	•	GGC Gly	710 GGC Gly
420	ACC CTC Thr Leu	4 TAT TYE		AGG Arg		GTG Val	610	ATC Ile	* 099	CTA Leu	AGT
	CAT His	TAC Tyr		CGC Arg	260	GCC Ala	9	AGT Ser		CCT Pro	ACG Thr
	() (I)	o GAC ASP	510	ACC Thr		ATG Met		CCA Pro		ACT Thr	700 TT GGA SP Gly
410	AGT	460 CAT GA		ACC Thr		GCA Ala		AAG Lys	029	GTG	GA
4	TCC	TGC		CGC Arg	0	GAG Glu	009 *	AAG Lys		GTG Val	CTT Leu
	GGA G	CCC	200:	ATT Ile	550	AGG		AAG Lys		$_{\rm GLY}^{\rm GGT}$	CTG
0	CGC (Arg (450 GAG (Glu)	Ŋ	CCC		TCC		ACA	640	ATG Met	690 AAT Asn
400	CTC (TTC		AGA Arg		CCT Pro	590	ACC Thr	9	GGA Gly	AAT Asn
	GCC	TGC Cys	0	TCC	540	TCC	u,	GTT Val		ACT	TAC
	TCC (440 GCC Ala	490	GGA Gly		GCT Ala		GAA Glu		GTG Val	680 TTC Phe
390	TCC	4 CTC Leu		TTC Phe		CGA Arg	0	CAG	630	GTT Val	GTT Val
	GCT Ala	TAC		$ ext{TTG}$	530	AAT Asn	580	GAA Glu		GTA Val	GAT Asp
	TCT (터보	480	* TCC Ser	S	CTC		CCT		CGA	670 GAC CCT ASP Pro
380	CCT 1 Pro 9	430 ACC TC Thr Se	7	GCA		AGG		CAA	620	CGG Arg	670 GAC CO ASP P:

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. 0	GCT Ala>	810	CTC Leu>		AAG Lys>		CTA Leu>	ATG Met>	00	AAG AAG Lys Lys>	1050	GCT Ala>
760	ATT Ile		AAG Lys		GGC G1y		GAG Glu	950 GGA Gly	1000	AAG Lys		TCA
	AGA		CCG	0	GCT Ala	006	AAA Lys	GGT G1y		\mathtt{TAT}		GGA Gly
	ACG Thr	800	GCC Ala	850	ACC Thr		ATG Met	ATG Met		TCA Ser	1040	ATG Met
750	CCT	8	GTG Val		CTG		GTG Val	0 GCA Ala	066	ATT Ile	1(AAT Asn
	TTT Phe		TGG Trp		ATG Met	068	GAT	94 TCA Ser		AGG Arg		ACA Thr
	CAA Gln	0	$_{\rm GGT}$	840	TAC Tyr	ω	GAA Glu	GGC Gly		GCC CTA Ala Leu	30	ACC Thr
740	GCT	790	GAT Asp		CTA Leu		ACC Thr	ATT Ile	086	GCC Ala	1030	GCT
7	TGT Cys		ACA Thr		ATG	880	ATC Ile	930 CTC Leu	01	GAA Glu		TTC
	GAT Asp		TCC Ser	830	TTC Phe	88	GGA Gly	GTT Val		ATT Ile		CCT
0	TTT Phe	780	TTC Phe	ω	AAG Lys		$_{\rm GGT}$	GGA Gly	970	GCC	1020	GTA Val
730	ACC Thr		TCT Ser		GAC Asp		GAT Asp	920 TGC Cys	9	GAT Asp	• •	TGT Cys
	GAG Glu		AAG Lys	0	ATG Met	870	ACA Thr	AAA Lys		AAT Asn		TTT Phe
	ATA Ile	170	ATC Ile	820	AGG Arg		TTA Leu	AGA Arg		TTC Phe	1010	CCC Pro
720	GAG Glu	7	GAG Glu		AAG Lys		GCA Ala	910 GAT AAA ASP LYS	096	GTA Val	1(AAT Asn
	AGC		GGA Gly		TCT	860	AAA Lys	91 GAT ASP		AAG Lys		ATG Met

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	TCT Ser>		CAT His>	GCG. Ala>	40	TTG Leu>	1290	AGT Ser>		CTA Leu>	•	GAA Glu>
	ATA Ile		AAC Asn	1190 A GAT er Asp	1240	GCT Ala		GAC		CTA		GCA Ala
0	TCG	1140	GCG Ala	11 TCA Ser		CGA Arg		TGG	30	CTA Leu	1380	TAC
1090	TAC TCG Tyr Ser	г	GCT	GGC Gly		TGC	1280	CCA	1330	GTG Val		ATT Ile
	AAC Asn		AAT Asn	30 GGG Gly	1230	GCA Ala	12	AGA Arg		GGA Gly		ACT
	CCC Pro	1130	ATG Met	1180 TGC GGG Cys Gly	1	GTT Val		TCA		GCT Ala	1370	GCG
1080	GGG G1y	11	ATA Ile	CTT		TTT Phe	0,	GCT Ala	1320	GGA G1y	H	AAA AGA GGT Lys Arg Gly
Т	ATG Met		TGT Cys	ATG Met	1220	GGT Gly	1270		` '	GAA Glu		AGA
•	TGG Trp	0	TTT Phe	1170 GAT GTG Asp Val	13	GGA Gly		ACT		${\tt GGG}$	09	AAA Lys
1070	GGA Gly	1120	AAC Asn	1 GAT ASP		ATG Met		CCT	1310	ATG Met	1360	AAG Lys
10	TTG		AGT Ser	GCA Ala	0;	GGT Gly	1260	GAC Asp	Ä	GTT Val		GCA Ala
	GAC Asp		ACG Thr	1160 3C GAA 3Y Glu	1210	ATT Ile		TCC		TTT Phe		CAT
0		1110	GCA Ala	1160 GGC GAA Gly Glu		CCT		AAT Asn	00	GGA Gly	1350	GAG
1060	GCA ATG Ala Met	7	TGT Cys	AGA Arg		ATA Ile	1250	AGA Arg	1300	GAT Asp		TTG Leu
	CTT		GCT Ala	30 ATC Ile	1200	ATC Ile	12	CAG Gln		CGT		GAG Glu
	ATG Met	1100	ACT Thr	1150 ATA AT Ile I	П	GTA Val		TCC		AAT Asn	1340	GAG Glu

FIGURE 7

Ile Arg Thr Gly Trp Ile His Pro Asn Ile Asn Leu Glu> Glu Leu Lys Val Asn Ser Thr Lys Ser Met> His Leu Leu Gly Ala Ala Gly Gly Val Glu Ala Val Ser Val> Thr Pro Ala Gly Asp Ile Lys Glu Tyr Gln Ala Leu Ile His> Ile Asn Ala His Ala> CAC CCT GAT GGA GCT GGA GTG ATT CTC TGC ATA GAG AAG GCT TTG GCT His Pro Asp Gly Ala Gly Val Ile Leu Cys Ile Glu Lys Ala Leu Ala> GTT CAG GCA ATA AGG ACT GGG TGG ATC CAT CCG AAT ATT TTG GAA ATT GGT CAC CTT CTC GGA GCA GCC GGT GGT GTG GAA GCA GTT TCA GTA Tyr His Met Thr Glu Pro> TGT TTC GGC CAA AAC AGA GAG TTA AAA GTT AAT TCA ACC AAA TCA ATG ACA TCC ACT CCG GCT GGA GAT ATC AAA GAG TAC CAA GCT CTT ATC CAC CAT GCC TAC CAC ATG ACC GAG CCT 1430 CAG TCA GGA GTC TCT AGG GAA GAC GTA AAT TAC ATA AAT GCC Gln Ser Gly Val Ser Arg Glu Asp Val Asn Tyr Ile Asn Ala 1520 1710 1660 သည Gly Ser Phe Thr Cys Asp Ala 1560 1510 ACT TGC GAT 1460 1650 1600 Gly Gln Asn Arg GGG AGT TTC 1500 1690 1450 1640 1590 Ala TTT CTA GGT Phe Leu Gly 1490 Ile Gly Val Gln Cys Phe 1440 1630 1580

FIGURE 7 5/7

1770	AAG AAG Lys Lys>		TTT GGT Phe Gly>	1870	GTTTCCGTGT	1930	GTTGGTAGCT	1990	GAACCATGAC	2050	GTAGAGCAAT	2110	GTTGTACTTT	2170	CACGTAGTAA
1760	GTG GGT CCT Val Gly Pro	1810	TCA TTT GGG Ser Phe Gly	1860	ATC TAG GAC Ile ***>	1920	AGTTTTGAGG ACTCCAGCAT	1980	CCATGAGTTT TGTGTCCGGA GCTTTAGTCG GAACCATGAC	2040	AGAATTGTTG	2100	CCTTGCAATA	2160	ATCGAGTCAG TGAAGAAGAG AACAAAGCTG TTAACTCGGG
1750	AAA TTG CTC (Lys Leu Leu	1800	TTG TCT AAT Leu Ser Asn	1850	GCC CCT TAC Ala Pro Tyr	1910	AGTTTTGAGG	1970	TGTGTCCGGA	2030	ACTCCTTGCT	2090	AAATCTCCCT	2150	AACAAAGCTG
	GAT ACA Asp Thr	1790	GTC GGT Val Gly	1840	CTC TTC Leu Phe	1900	TCAAAGCTGA	1960	CCATGAGTTT	2020	CACTTGATAT	2080	TTTTTCTCTG	2140	TGAAGAAGAG
1740	GAA GGC GTG Glu Gly Val		AAC GTT AAG Asn Val Lys	1830	TCG TCC ATA Ser Ser Ile	1890	GTGGAATTCT ACTCAACATA TCAAAGCTGA	1950	CTAGACATGC	2010	CTCATGGCGA	2070	TCATATTTTT	2130	ATCGAGTCAG
1730	AAC CCA GAT Asn Pro Asp	1780	GAG AGA CTG Glu Arg Leu	820	GGG CAC AAC Gly His Asn	1880	GTGGAATTCT	1940	CCTTACGTCT	2000	GGATTGAGTA	2060	ATTCATTATC	2120	CGAGCTTTTC

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7	
FIGURE	7 / 7

AAAAAAAAA	TGGAAATAAA AAAAAAAA AAAAAAAA AAAAAAAA AAAAAA	AAAAAAAAA	AAAAAAAAA	AAAAAAAAA	TGGAAATAAA
2350	2340	2330	2320	2310	2300
ATGTATGTTT	AACTAGAAGA CTGGTTTAGA TTGGTTTGTT TTCTCATTGA TAATTGGGGR ATGTATGTTT	TTCTCATTGA	TTGGTTTGTT	CTGGTTTAGA	AACTAGAAGA
2290	2280	2270	2260	2250	2240
AAATTTGTAA	CCATTIGCCC TITGITITGC ICTCIATITC ATCACCGITT IGTGGTTTTA AAATTIGTAA	ATCACCGTTT	TCTCTATTTC	TTTGTTTTGC	CCATTTGCCC
2230	2220	2210	2200	2190	2180

2360 AGGGGGCCG CTCTAGAGG

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09	CACACCAAAC	120	ACAGACAGAC	180	TCTTCGATTC	240	TCCCAAAGGG	300	CCTGCCGCCT	360	TCTACCTCCT	420	CTCTCCCGCC	4.80	CTCCCAATGC GCCCCACTAC CTTCTGCTTC CTCCGCCCTC CGCGGATCCA
20		110	AGACAGACAG	170	CCTCCTTTCA	230	GGGTCTTTCA	290		350	CGCCTGCATG	410	TCGCCGACGC	470	CTCCGCCCTC
40	ACGCGTCCGC	. 100	CATTGGCAGC	160	ATGCGGCCAC	220	CGCCTTTTCC	280	TCAGTCAGTT	340	GGCTCCTTGC	400	TCTCCTCTCC	460	CTTCTGCTTC
30	GGGTCGACCC	06	AGACGGACGC	150	GATCCATCGA	21.0		270	TCTCAAAGGG	330		390	CCGCCTTCCA	450	GCCCCACTAC
20	CGGAATTCCC	80		140	GAGAGAGAGG	200	CATTCCGCTG	260	TATCCTATCT	320	CGCTTCCCCT	380		440	CTCCCAATGC
. 10	-A-CNTGGTC	. 70	TTCCTCAGCT	130	CCATAAAAGA	190	ATTACCATAC	250	TATCCTTTTC	310	CTTCCCTGCT	370	TCCACCCTC	430	GCCGGATTCT
	20 30 40 50	20 30 40 50 CGGAATTCCC GGGTCGACC ACGCGTCCGC GACGCCAACC CACACCAA	20 30 40 50 CGGAATTCCC GGGTCGACCC ACGCGTCCGC GACGCCAACC CACACCA 80 90 110	20 30 40 50 CGGAATTCCC GGGTCGACCC ACGCGTCCGC GACGCCAACC 80 90 100 110 TCTCTTCTCA AGACGGACGC CATTGGCAGC AGACAGACAG	20 30 40 50 CGGAATTCCC GCGTCGACC ACGCGTCCGC CACACCAACC 80 90 100 110 TCTCTTCTC AGACGGACG CATTGGCAGC AGACAGACA 140 150 160 170	20 30 40 50 CGGAATTCCC GGGTCGACCC ACGCGTCCGC GACGCCAACC R0 90 100 110 TCTCTTCTC AGACGACG CATTGGCAGC AGACAGACAG GAGAGAGAG GATCCATCGA ATGCGGCCAC CCTCCTTTCA	20 30 40 50 CGGAATTCCC ACGCTCCGC ACGCTCCAACC CACACCAACC 80 90 110 110 TCTCTTCTCTC AGACGGACG CATTGGCAG AGACGACA TCTCTTTCTC AGACGGACG ATTGGCAG ACAGACA GAGAGAGAG GATCCATCGA ATGCGGCCAC CCTCCTTTCA CAGAGAGAG ATGCGGCCAC CCTCCTTTCA TCTTCGA AGAGAGAGAG ATGCGGCCAC CCTCCTTTTCA TCTTCGA	20 30 40 50 CGGAATTCCC GGGTCGACCC ACGCGTCCG GACGCCAACC 10 100 110 TCTCTTCTC AGACGACG CATTGGCAGC GAGAGAGAG CATTCGCCCAC CCTCCTTTCA GAGAGAGAG GATCCATCG CCTCCTTTCA CATTCCGCTG ATCCATTTCC GGGTCTTTCA	20 30 40 50 CGGAATTCCC GGGTCGACC ACGCGTCCGC CACGCTCCC 80 90 100 110 TCTCTTCTC AGACGGACG CATTGGCAGC ACAGGACA GAGAGAGG CATTGGCAGC ACAGGACA ACAGGACA GAGAGAGG GATCCATCGA ATGCGGCCAC CCTCCTTTCA TCTTCGA CATTCCGCTG ATGCGGCCAC CCTCCTTTCA TCTTCGA CATTCCGCTG ATCCATTTCC GGGTCTTTCA TCCCAAA CATTCCGCTG ATCCATTTCC GGGTCTTTCA TCCCAAAA	20 30 40 50 CGGAATTCCC GGGTCGACCC ACGCGTCCGC CACACCAACC 80 90 110 110 TCTCTTCTC AGACGGACG CATTGGCAG AGACGACAG ACAGACAG TCTCTTCTC AGACGGACG CATTGGCAG ACAGACAG ACAGACAG GAGAGAGG GATCCATTGG ATGCGGCCAC CCTCCTTTCA TCTTCGA CATTCCGCTG ATCCATTTC GGGTCTTTCA TCCCAAA TATCCTATCT TCTCAAAAGGG TCAGTCAATG CCTCCCAATG TATCCTATCT TCTCAAAAGGG TCAGTCAATG CCTCCCAAATG	20 30 40 50 CGGAATTCCC ACGCGTCCGC GACGCCAACC CACACCAACC 80 90 100 110 ACACACCAACC TCTCTTCTCA AGACGGACGC CATTGGCAGC ACAGACAACA ACAGACAACA GAGAGAGAG CATTGGCACC CCTCCTTTCA TCTTCGA CATTCCGCTG ATGCGCCCAC CCTCCTTTCA TCTTCGA CATTCCGCTG ATCCATTTC GGGTCTTTCA TCTCCAAA CATTCCGCTG ATCCATTTC GGGTCTTTCA TCTCCAAA TATCCTATCT ATCCAAAGGG TCAGTCAGTT CCTCCCAATG CCTGCCGGGCCGG TATCCTATCT TCTCAAAGGG TCAGTCAGTT CCTCCCAATG CCTGCCGGGG CCTGCCGGGGGGGG TATCCTATCT TCTCAAAAGGG TCAGTCAGTT CCTCCCAATG CCTGCCGGGGGGGG CCTGCCGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	20 30 40 50 CGGAATTCCC ACGCGTCCGC GACGCCAACC CACACCAACC ROBATTCCC ACGCGTCCGC GACGCCAACC CACACCAACC TCTCTTCTCA AGACGGACGC ACACGACAC ACACGACAC GAGAGAGAG ATGCGGCCAC CCTCCTTTCA TCTTCGA CATTCCTCCCT ATCCATTTC CCTCCTTTCA TCTCCAAA CATTCCTATTC ATCCAAA ACCCTTTTCA TCTCCAAA TATCCTATCT TCTCAAAAGG TCACTCCAATG CCTCCCAATG TATCCTATCT TCTCAAAAGG TCACTCCAATG CCTCCCAATG TATCCTATCC TCTCTATACG TCTCCTATTC CCTCCCAATG	20 30 40 50 CGGAATTCCC ACGCGTCCGC ACGCGTCCGC CACGCTCCGC CACGCTCCGC CACGCTCCGC CACGCTCCGC CACGCTCCGC CACGCTCCGC CACGCTCCGC ACGCGACGC ACGCGACGC ACGCGACGC ACGGACGACG ACGGACGACG ACGGACGACG ACGGACGCC ACGGACGACG ACGCACGACG ACGCACGACGACG ACGCACGACGACG ACGCACGACGACG ACGCACGACGACG ACGCACGACGAC	20 30 40 50 CGGAATTCCC GGGTCGACC ACGCGTCCG GACGCCAACC R 90 110 111 TCTCTTCTCA AGACGGACG ATGCGCCAC AGACGACAG GAGAGAGAG ATGCGCCCAC CCTCCTTTCA 230 CATTCCTATT CACTTTTCA 220 230 TATCCTATT CGCCTTTTCA 320 320 TATCCTATC TCTCAAAGG TCACTCCAATG CCTCCCAATG TATCCTATCC TCTCTATACGT CCCTCCAATG TATCCTTCCC TCTCTTACG GGCTTCCAATG CGCTTCCCTT CCCTGCATGC CGACCTTCCT TCTCTTTCC S80 400 410 CGACCCTTCCT TCTCCTTCTC	20 30 40 50 CGGAATTCCC ACGCTCCGC GACGCCAACC CACACCAACC 80 90 100 110 TCTCTTTCTC AGACGGACG CATTGGCAGC ACACAGACAG ACAGACAC GAGAGAGAG ATCCATTTC ATCCATTTC TCTTCGA TCTTCGA CATTCCGCT ATCCATTTTC GCCTTTTCA TCTTCGAA TATCCTATC ATCCATTTCC GGTCCTTTCA TCTCCAAA TATCCTATC TCTCAAAGG TCTCCAAA TCTCCAAA TATCCTATC TCTCTAAGG TCTCCAAAGGG TCTCCCAAAGGG TCTCCCAAAGGG TATCCTATC TCTCTTTCA TCTCCCAAAGGG TCTCCCAAAGGG TCTCCCAAAGGGG TCTCCCAAAGGGG TATCCTCCC TCTCTTTCC TCTCCCAATGG TCTCCCGGGCCG TCTCCCGGCG CGCTTCCCCT TCTCTCTTCC TCTCCCCAAGGGG TCTCCCCGGCCG TCTCCCCGCG CGCTTCCCAATGG TCTCCCCAACGGC TCTCCCCCGCCCG TCTCCCCCGC TCTCCCCCGC ABB ABB ABB ABB TCTCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC

FIGURE 8 1/5

540	GACTACTATA	* 009	CGGAGGCTCA	* 099	ACAGGAAGTT	720	AATGGGTGTG	780	ATGGAACGAG	840	TTGCTGGAGA	*	GGATGGACAA	* 096	GAATCACCGA	1020	CTCATTGGCT CAGCAATGGG
530	GCCCTGCCAT	290	CCGCAGGCAC	650	TGCAACCTGA	710	TTGTGACTGG	770	AATCTGCTTG	830	CCTACGAGAA	890	CTCTCTAAGA	950	GAAAGCATTA ACAGATGGTG	1010	
520	CCTGCTTCGA GCCCTGCCAT	580	TTCGCACCAC	640	CCCTTCCAGG GGAGGCAATG GCCGTGGCTC TGCAACCTGA ACAGGAAGTT	700	CGGCGAGTAG	760	TTTCTACAAT	820	TGCTCAATTT.	880	GGCCCCGAAG	940	GAAAGCATTA	1000	ATGCGGAGTT
510	TCTTACCTCG	570	TCCAGACCCA	630	GGAGGCAATG	069	TATCAAACAG	750	ACCTGATGTT	810	CCTTTGATTG	870	ATGGTTGGGT	930	TACATGCTGA CTGCTGGCAA	066	AAAGAGCTAG ATAAAAGAAA
200	CCTCGTCACC	260	CTTGTTCGGA TCCAGACCCA TTCGCACCAC	620	CCCTTCCAGG	089	ACCACAAAGA AGAAGCCAAG	740	TAGGCCATGA	800	GAGATAGAGA	860	GATCAAGTCT TTCTCCACAG ATGGTTGGGT	920		086	
490	GTTTCCATAC	550	CATCCGCATC	610	ATCGAGCTTC	0.19	ACCACAAAGA	730	GTGACTCCTC	790	TGGCATAAGC	850	GATCAAGTCT	910	GTTCATGCTA	970	AGATGTGATG

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ACCACATGAC	GAGTTTCACT TGCGATGCCT ACCACATGAC	GAGTTTCACT	TGCGACTATT TACGCAGAAT TTCTAGGTGG	TACGCAGAAT	TGCGACTATT
1500	1490	1480	1470	1460	1450
AGAAAAGAGG	GAGCATGCAA	AGAGGAGTTG	TATGGGGGAA GGAGCTGGAG TGCTACTACT	GGAGCTGGAG	TATGGGGGAA
1440	1430	1420	1410	1400	1390
ATGGATTTGT	AGTAATCGTG	GAGAAATTCC GACCCTACTA AAGCTTCAAG ACCATGGGAC AGTAATCGTG	AAGCTTCAAG	GACCCTACTA	GAGAAATTCC
1380	1370	1360	1350	1340	1330
CTTTGTCCCA	GCATGCCGAG	TIGGTATGGG AGGITTTGIT GCATGCCGAG		ATCATACCTA	AGATGCGGTA
1320	.1310	1300	1290	1280	1270
GCGGGGCTC	GTGATGCTTT	CGAAGCAGAT	TAATCAGAGG	GCGAACCATA	AATGAATGCT
1260	1250	1240	1230	1220	1210
ACTTTTGTAT	GCAACGAGTA ACTTTTGTAT	TACTGCTTGT	GGGATGGATG GGGCCCAACT ACTCGATATC TACTGCTTGT	GGGCCCAACT	GGGATGGATG
1200	1190	1180	1170	1160	1150
CAATGGACTT		TCCCTTTTGT GTACCTTTCG CTACCACAAA TATGGGATCA GCTATGCTTG	CTACCACAAA	GTACCTTTCG	TCCCTTTTGT
1140	1130	1120	1110	1100	1090
AGAAGATGAA	ATTTCATATA	GTATTCAATG ATGCCATTGA AGCCCTAAGG ATTTCATATA AGAAGATGAA	ATGCCATTGA	GTATTCAATG	TGGAATGAAG
1080	1070	1060	1050	1040	1030

FIGURE 8

1560	TGGCTCAGTC	1620	CICCGGCIGG	1680	AGTTAAAAGT	1740	TGGAAGCAGT	1800	TGGAAAACCC	1860	TGAACGTTAA	1920	TCTTCGCCCC	1980	GAAGTTTTGA	2040	TTTGTGTCCG
1550	GAGAAGGCTT TGGCTCAGTC	1610	GCCACATCCA	1670	CAAAACAGAG	1730	TTGGTCACCT TCTCGGAGCA GCCGGTGGTG	1790	AATATTAATT	1.850	AAGGAGAGAC	1910	TCGTCCATAC	1970	TATCAAAGCT	2030	CTCCTTACGT CTCTAGACAT GCCCATGAGT
1540	TCTCTGCATA	1600	AAATGCCCAT	1660	CTGTTTCGGC	1720	TCTCGGAGCA	1780	GATCCATCCG	1840	GGGTCCTAÀG	1900	TGGGCACAAC	1960	CTACTCAACA	2020	CTCTAGACAT
1530	CTGGAGTGAT	1590		1650	CTCTTATCCA	1710	TTGGTCACCT	1770	CAGGCAATAA GGACTGGGTG	1830	GTGGATACAA AATTGCTCGT	1890	TTGGGTTTGG	1950	GTGTGGAATT	2010	CTCCTTACGT
1520	CCTGATGGAG	1580	AGGAGTCTCT AGGGAAGACG TAAATTACAT	1640	GAGTACCAAG CTCTTATCCA CTGTTTCGGC	1700	AAATCAATGA	1760	CAGGCAATAA	1820		1880	TCTAATTCAT	1940	GACGTTTCGT	2000	GGACTCCAGC ATGTTGGTAG
1510	CGAGCCTCAC	1570	AGGAGTCTCT	1630	AGATATCAAA	1690	TAATTCAACC	1750	TTCAGTAGTT	1810	AGATGAAGGC	1870	GGTCGGTTTG	1930	TTACATCTAG	1990	GGACTCCAGC

FIGURE 8

		ATCC	2370 GCTCTAGAGG	2350 2350 2360	2350 AAAAAAAAA
TTTTCTCAAA	AAAACTAGAA GACTGGTTTA GATTGGTTTG	GACTGGTTTA	AAAACTAGAA	TAAAATTTGT	TTTGTGGTTT
2340	2330	2320	2310	2300	2290
TCATCACCGT	GCTCTCTATT	CCTTTGTTTT	TGTTAACTCG GGCACGTAGT AACCATTTGC CCTTTGTTTT	GGCACGTAGT	TGTTAACTCG
2280	2270	2260	2250	2240	2230
AGAACAAAGC	AGTGAAGAAG	TCATCGAGTC	CTCCTTGCAA TAGTTGTACT TTCGAGCTTT	TAGTTGTACT	CTCCTTGCAA
2220	2210	2200	2190	2180	2170
TGAAATCTCC	TTTTTTTCTC	TCTCATATTT	CTAGAATTGT TGGTAGAGCA ATATTCATTA TCTCATATTT TTTTTTTCTC	TGGTAGAGCA	CTAGAATTGT
2160	2150	2140	2130	2120	2110
ATACTCCTTG	GACACTTGAT	TACTCATGGC	ACGGATTGAG TACTCATGGC GACACTTGAT ATACTCCTTG	CGGAACCATG	GAGCTTTAGT
2100	2090	2080	2070	2060	2.050

7.1GUKE 8

Sequence Range: 1 to 1580

GGG Gly>	0	TCG Ser>	150	GTG Val>		GAT Asp>		TCT Ser>	AAA Lys>	340	CGC Arg>
50 TCT Ser	100	CAT His		AGG Arg		GGT Gly		GGA Gly	290 GCT	ř	ATC Ile
GCA Ala		CAG		AAA Lys	0	TTG Leu	240	ATT Ile	CTT Leu		${\tt GGG} \\ {\tt Gly}$
AAT (140	TCC	190	TCT Ser		TTA Leu	GAT Asp		ACG Thr
	06	GCA ACT Ala Thr	-	GTC Val		CAG Gln		AAA Lys	280 AAT GAT ASn ASP	330	CGA Arg
40 ATG GCG Met Ala		AGG Arg		TTT Phe		AGG Arg	230	TGC Cys			GTC Val
366		AGA Arg	0	GAG Glu	180	GAC	(7	GGA Gly	TCA Ser		ACT Thr
GCT	80	CTG	130	TCG		TCT Ser		AGA Arg	GTC Val	320	ATT Ile
30 CGTT		GCC		TCC		GAT Asp	0	AGT Ser	270 CAA Gln	.,	TGG Trp
GTTT(CCT		TCT Ser	170	CAG Gln	220	GTG Val	CTT Leu		GAA Glu
O A GA	0	GTT Val	120	GGA Gly	\leftarrow	GTT Val		CTT Leu	GCT Ala	310	GAT Asp
20 AGAGA	70	TCA		CGT Arg		GCC Ala		AGG Arg	cca Pro	31	AAT Asn
ľTCA.		TCT		TCT Ser	0	AGT Ser	210	CCG Pro	2 ATA Ile		ACC Thr
10 GG A		GGT Gly	110	TCG	160	TGT Cys		TCG	GCT Ala		GAC Asp
AATC	09	CTG (TCA		TGC Cys		CGC Arg	O TCT Ser	300	GTC Val
10 CCTGAATCGG ATTCAAGAGA GAGTTTCGTT GCTGGG		TTT (Phe]		ATT		TTT	200	TCT	250 GGT TCT Gly Ser		ATT Ile

FIGURE 9 1/5

390	TCA Ser>		GAT Asp>		GGC Gly>	TTG Leu>	280	GTC Val>	630	GTG Val>		GGA Gly>
, /	GCA A		AAT Asn		TTC Phe	530 CCT Pro	28	TTA Leu		CTA Leu		CGG Arg
	TTA (Leu 1	0	GCA	480	CTT Leu	5 AAT Asn		GGT Gly		ATT Ile	019	GAT Asp
380	AAT ?	430	GAC (Asp		GAC	AAG Lys		TTG	620	AAT Asn	9	ACC Thr
38	ACA 7		GTA (Val		70 =	A 10	570	GTG Val	9	AAC Asn		TGG Trp
	CTT /		CAG (Glu	470	CCT Pro	520 TGC AAA Cys Lys		TTT Phe		TTT Phe		GAC
0	AGT (420	GCA Ala	4	ACC	GGC Gly		GGA Gly	610	GGT Gly	099	GTT Val
370	GAT Asp		ATG		TCT Ser	CTT	260	AGT Ser	61	GGG Gly		\mathtt{TAT}
,	AAA (Lys		GAG Glu	0	ACT Thr	510 GCA Ala	Lr)	TGC		GGT Gly		CGG Arg
	GGT	410	CTA	460	TGT Cys	AAA Lys		GCA Ala		AGA Arg	650	CTT TCT Leu Ser
360	TCA	4	GCT		ATG Met	TCG Ser	550	GCT Ala	009	ATT Ile		
	CTC		AAA Lys		TTG	500 ATA Ile	5.	ACC Thr		CAC His		TCT
	GTT Val	0	AGG	450	GTT Val	5 CAG Gln		ATT Ile		TGC	640	GAT Asp
350	AGG	400	GCA		ATG Met	CCT	•	GAC Asp	590	GCT Ala	9	GCT
т	CGA Arg		GCA Ala		GAT Asp	90 GCT Ala	540	TAC TYr	Ξ,	GCT Ala		GGT Gly
	AAC Asn		GAG Glu	440	GTG Val	490 AGT G		. TCT Ser		TCA		ATT Ile

FIGURE 9
2/5

720

710

700

Trp Leu Leu Leu His Gln Ala> Ile Glu Ser Ala Leu Gly Lys> GGA GAT GGG CAA AGG CAT CTA AAA GCT GCA ATC AAA GAA GAT GAA GTT Gly Asp Gly Gln Arg His Leu Lys Ala Ala Ile Lys Glu Asp Glu Val> Glu Val Phe Arg> Ile Arg Asp Phe Pro Pro Arg> 990 1010 TCC AAC ATC GAC TGG TTG CTG CTT CAT CAG GCA GGA GCT GTA GTG GTG CAG TCA Gly Ala Val Val Val Gln Ser> Leu Phe Ala Phe Asp Leu His Ser Asp> 870 CTT GGA AAG CCA AGG GAG GTA TTC CGC TTT GAT TTG CAT AGC GAT TCC ATC AGA GAT TTT CCA 910 CCT CAG TCA ATC GAA TCA GCA 860 TGC ATC CAA ATG AAC GGT AAA Cys Ile Gln Met Asn Gly Lys 810 1000 950 GCT 006 850 Pro Gln Ser Ile Asp 750 CTC TTT (Ala (GAT AAA GCC CTG GGA CAT AAT GGG TCC Asp Lys Ala Leu Gly His Asn Gly Ser 800 Asp Ala GCT940 Ser Asn Glu Glu Asp Gly GAT GAG GAA GAT GGG 890 CGC TCT GTG (Arg Ser Val F Thr Cys Ile Leu Phe Gly GGA 840 790 TCT Ser GCC GGT CTT AAT GGA Ala Gly Leu Asn Gly TyrTCT TCA TAC 930 CTC Ser Cys TTT GCT TGC TGT GAT GCT Cys Asp Ala TGT ATT 830 Ala Arg Ser 780 970 Phe i ACA ' 920 680

FIGURE 9 3/5

1060	CCT CAA Pro Gln>	1110	GCG GCA Ala Ala>		GTG AAG Val Lys>		ACA TGG Thr Trp>	1260	CACTGCAGCT	1320	AAGAAGTCAG	1380	TCGTTCCCCT	
1050	CTA GAG GTT Leu Glu Val	1100	AAC ACT AGT Asn Thr Ser	1150	AGT GGA AAT Ser Gly Asn	1200	GCC GGA CTC Ala Gly Leu	1250	GCCGAGCCAG	1310	GCTTCCATGA CCANAAAAAG	1370	TTGCCCTTTT	
1(ACA CGT Thr Arg	06	TAC GGG Tyr Gly	1140	GTG AGG Val Arg	1190	TTT GGC Phe Gly	1240	GACTGAA	1300	CTTCCATGA	1360	CTTCATCACA	
1040	GCA GTA GCA Ala Val Ala	1090	TTG GCA AAT Leu Ala Asn		GAC GAA GCT Asp Glu Ala	1180	ACC GCA GGA Thr Ala Gly	1230	TGG GGA TAA (Trp Gly ***>	1290	BAAATTTT GO	1350		
1030	ATT GAT Ile Asp	1080	TCA AAC Ser Asn	1130	TTG GCA CTA G Leu Ala Leu A		CAC GTG ATT GCA A His Val Ile Ala T	1220	ATT ATC AGG TILE ILE AKG T	1280	CCGATGTTTC ACGAAATTTT	1340	AGCAAGCAAC ACGACACGAT	
1020	AAT CAG AGG ATC Asn Gln Arg Ile	1070	GAA CGA ATT ATC Glu Arg Ile Ile	1120	TCC ATT CCC TI Ser Ile Pro Le	1160 1170	CCG GGT CAC G' Pro Gly His Va	1210	GGT TCT GCT A' Gly Ser Ala I	1270	TCCTCTCAAA CC	1330	TCTTTTATGG AG	
						7								

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1440	TTGTCCCCAA	1500	CGGGACATTG	1560	AAAAAAAA				
1430	ATAGTTTCTT	1490	CATTTTGTCT	1550	AAAAAAAAA				
1420	TACAATACCC	1480	GCTTTTACTT	1540	TTTGCTAAAA				
1410	TTGCTGACAA	1470	TAATTGTTCA	1530	ATGTTTATAT				
1400	TTTGATGATT	1460	TAAGTTATTT GTTTCTTGTT TAATTGTTCA GCTTTTACTT CATTTTGTCT CGGGACATTG	1520	GAGATGACAG CATAAACATC ATGTTTATAT TTTGCTAAAA AAAAAAAAA AAAAAAAAA	1580 AAAAAAAAA			
1390	TITCCATTAG ITTGATGATT TTGCTGACAA TACAATACCC ATAGITTCTT ITGTCCCCAA	1450	TAAGTTATTT	1510	GAGATGACAG	1570 AAAAAAAA AAAAAAAAA			

FIGURE 9 5/5

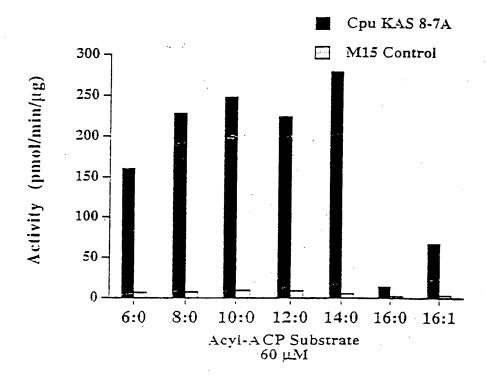


FIGURE 10

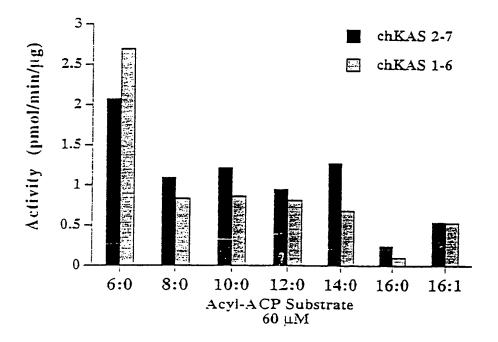


FIGURE 11

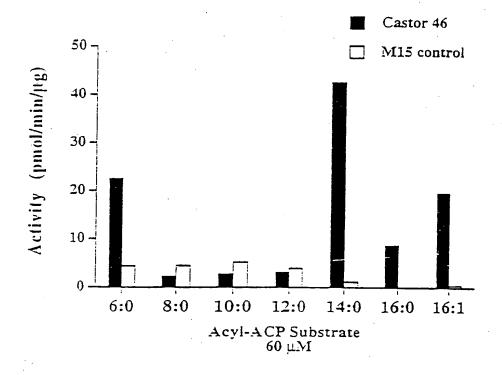
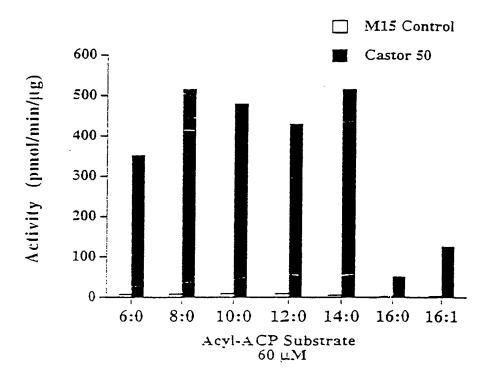


FIGURE 12



E328013-28

FIGURE 13

Cp FatB1 VS Cp FatB1/Kas A-2-7

Cp FatB1 VS Cp FatB1/Kas A-2-7

4 Mole % C:8

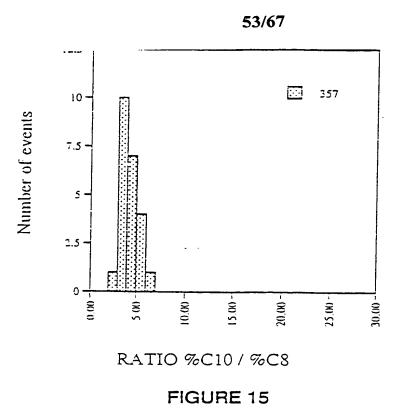
2

3

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7

FIGURE 14



1/2

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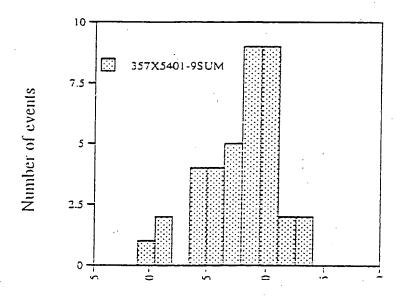
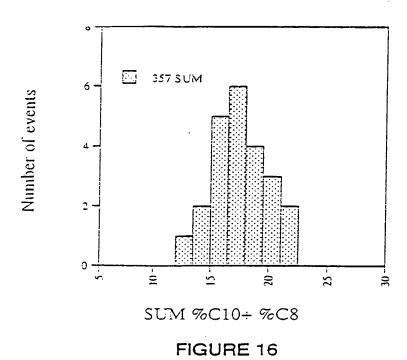


FIGURE 15 2/2



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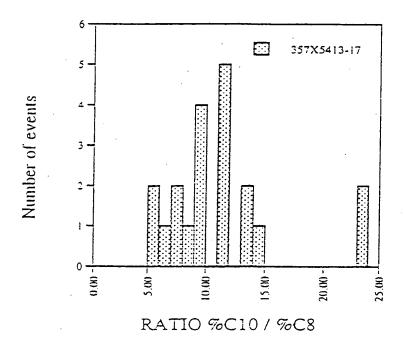


FIGURE 17

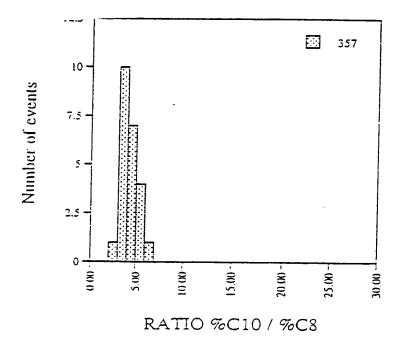


FIGURE 17

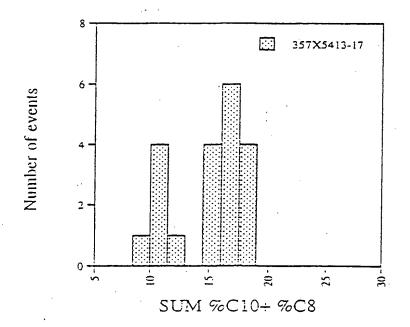
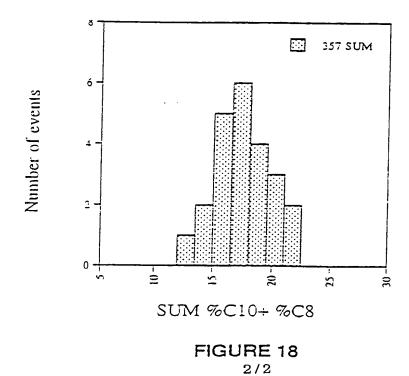
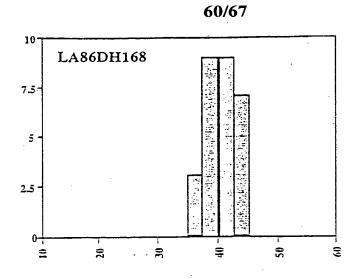


FIGURE 18



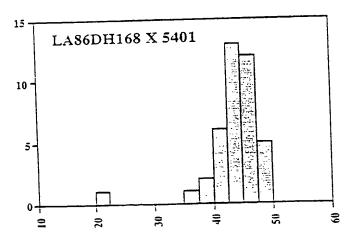
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12:0 levels (w%)

FIGURE 19 1/3



12:0 levels (w%)

FIGURE 19 2/3

Number of independent events

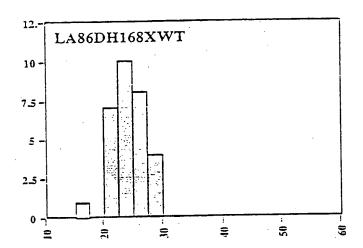


FIGURE 19 3/3

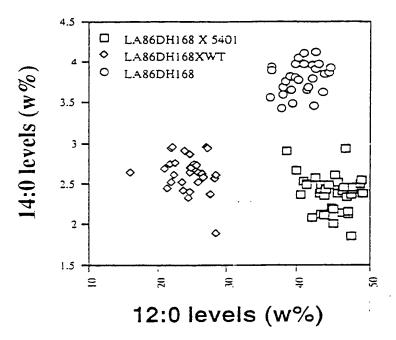
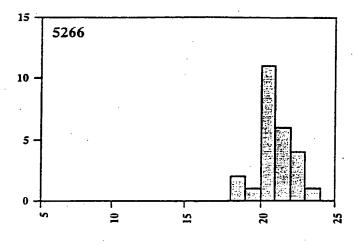


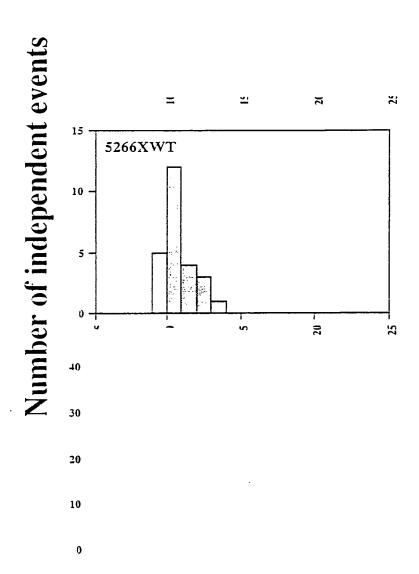
FIGURE 20

Number of independent events



18:0 levels (w%)

..FIGURE ~21



18:0 levels (w%)

FIGURE 21 2/3

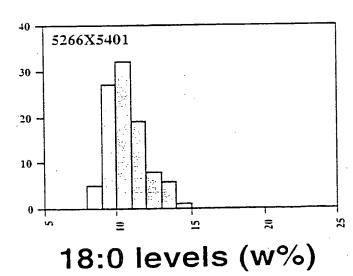


FIGURE 21 3/3

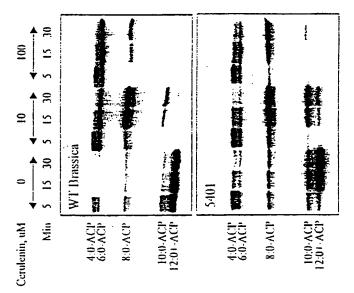


FIGURE 22

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International application No.

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Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically: see FURTHER INFORMATION sheet
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Claims Nos.: 1-14,19,20,21,26,27,28

Remark : Claims 1-14 were not provided to the ISA at the time of search and hence the subject matter of these claims and the dependent claims 19,20,21,26,27,28, could not be defined.

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